EFFECT OF IRON OVERLOAD ON 7, 12-DIMETHYL-BENZ (A) ANTHRACENE-INDUCED SKIN TUMORIGENESIS

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ABSTRACT

Iron overload is known to occur in the West European and American population due to the consumption of iron-rich diets. On the other hand, genetic disorders leading to iron overload are also known. Iron overload leads to increased peroxidation and disruptive disintegration of lipid-rich membranes, and predisposes humans for an enhanced risk of cancer induction. In experimental animals iron overload enhances intestinal, colonic, hepatic, pulmonary and mammary carcinogenesis. In this study, we have shown that iron overload is a mild tumor promotor in mouse skin. Female albino Swiss mice were iron overloaded and their backs were shaved. Tumors were initiated using a complete tumorigenesis protocol by applying 200μg 7,12-dimethylbenz (a) anthracene (DMBA)/mouse in multiple doses of 40μg DMBA/day for 5 consecutive days. The appearance of the first tumor (latency period), percent of tumor incidence and number of tumors/mouse were recorded. When compared to the positive control group, the iron overloaded mice showed an increased incidence of tumors. In iron overloaded animals, the tumors appeared about four weeks earlier. The number of tumors per mouse were significantly higher in the iron overloaded group. Biochemical studies performed in the present study include the determination of the activity of lipid peroxidation, catalase and xanthine oxidase measurement in mice skin tissue. We observed an iron-mediated induction in lipid peroxidation (LPO) and xanthine oxidase (XOD) and diminished catalase (CAT) activity in skin tissues of mice overloaded with iron as compared to the normal unloaded control group. Based on these studies we propose that iron increases tumor promotion potentials significantly. An induction in LPO in the iron overloaded group suggests that oxidative stress may be responsible for such an observed augmentation of cutaneous carcinogenesis in mice. Our data indicate that iron overload exerts tumor promoting potential in mouse skin, and that oxidative stress generated by iron overload is responsible for the augmentation of cutaneous tumorigenesis.


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INTRODUCTION

In developing countries, iron deficiency is an important nutritional problem. In developed meat eating countries, however, iron excess may be more of a problem than iron deficiency. Iron overload has been observed in the human population throughout the world. It has been estimated that 0.2 to 0.5% of humans have a genetically determined form of iron loading called hereditary hemochromatosis. In addition, iron overload has also been observed in patients with thalassemia major. In sub-saharan African countries, clinically and pathologically significant iron overload can result from the ingestion of home brewed beer which is rich in iron. Porphyria cutanea tarda, chronic liver diseases and chronic renal failure result in patients with significant degrees of iron overload. Epidemiological studies suggest that increased body iron stores may be associated with an increased risk of cancer and increased overall death rates. In experimental animals increased body iron has been shown to enhance mammary, colonic, intestinal and hepatic carcinogenesis. Exposure to iron-containing ores increases the risk of lung cancer and an increased incidence of lung cancer has been observed among smokers having high iron overload. These studies suggest that iron overload conditions may prime tissues for an enhanced incidence of carcinogenesis. However, to the best of our knowledge, no evidence has been provided to show that iron overload augments skin carcinogenesis. In this study, we provide evidence that iron overload acts as a cutaneous tumor promoter and enhances DMBA-induced skin carcinogenesis.

MATERIALS AND METHODS

Chemicals
Iron dextran (imferon) was purchased from Rallis India, Ltd. Hair removing cream was from Anne French, Geoffrey Manners and Co. Ltd. (India). 7,12-dimethylbenz(a)anthracene (DMBA), and metal iron in powder or crystal form was obtained from Sigma Chemical Co. (St. Louis, MO, USA). Three standard reference materials have been obtained from the International Atomic Energy Agency (Vienna, Austria) to be used as reference for iron estimations. These include 1) Mixed Human Diet, H-9, 2) Rye Flour, V-8, and 3) Hay powder, V-10. Nitric acid, perchloric acid and sulfuric acid of “analar” grade were procured from S.D. Fine Chemicals, E. Merck, and Glaxo (India), respectively. All other chemicals and biochemicals used in this study were either of analytical grade or of highest purity grade available commercially.

Animals
Swiss albino mice free from pathogen were procured from the Central Animal House Facility of Hamdard University and used throughout this study for our experiments. Animals were housed in an air conditioned room in polypropylene cages. They had free access to pellet diet (Lipton, India, Ltd.) and water ad libitum. For long term chronic experiments, usually 20 mice were housed and for acute experiments, six mice were taken in each group. The animals were kept at a room temperature of 24°C (±2°C) and were exposed to alternate cycles of 12 hour periods of light and darkness. The dorsal skin of the mouse was shaved with electric clippers (Oster A4) followed by the application of hair removing cream at least two days prior to the treatments. Only mice that did not show signs of hair re-growth were used.

Preparation of iron dextran solution
Each 10 mL dose vial containing 500 mg of iron as iron complex with a low molecular weight dextran fraction was diluted in saline to obtain a suitable dose.

Iron estimation conditions
The atomic absorption spectrophotometer gives results directly in ppm. All instructions for instrument setting, calibration and assay as laid down in the manual were strictly followed.

Serum tissue iron estimation
For serum preparation, blood was collected from the retro-orbital sinus by heparinized capillary tubes into dry test tubes which were kept in tilted positions for 10 minutes at room temperature and then for an hour at 4°C in order to separate the serum from the cellular clot. To remove cellular contamination from the serum, it was centrifuged for 2-3 minutes at 800 x g. After collection of blood serum, 0.5 mL of serum was transferred to a glass vial containing 1 mL of digestion mixture (1 part of perchloric acid mixed with 3 parts of conc. nitric acid and left for 3 hours at room temperature). It was left overnight, then the glass vials were heated to near dryness on a hot plate. 5 mL distilled water was added to it after which it was slightly heated up to 40-50°C and then filtered using Whatman filter paper through a Buchner funnel into a test tube. The filtrate was read on ASS. Serum iron concentration was calculated and represented as parts per million or µg/mL serum. Tissues such as liver and skin were excised, cleaned free of extraneous material and washed immediately with saline (0.9%). In case of skin, the dorsal skin of mice was removed, scraped off with a surgical blade to remove fat and blood vessels and was used for iron estimation. 1.0 g of the tissue was taken in a glass vial and added to 10 mL of concentrated HNO3. The samples were left overnight. The next day, the samples were found to be dissolved in the acid to a large extent. They were then dried on a hot plate to near dryness. 5 mL
Estimation of tissue iron level

For estimating tissue iron levels in normal and iron overload states, blood samples were taken and divided into two groups. Group I received an intramuscular injection of saline whereas group II received an intramuscular injection of imferon (5 mg iron/0.2 mL saline/mouse) daily on the dorsal skin 24 hours following the last injection of saline or imferon. No further treatment in any of the two groups of mice was given. The mice were observed weekly for the occurrence of tumors, which were recorded and plotted as function of weeks on test. The data were presented both as number of tumor per mouse as well as percent incidence of tumorigenesis.

Diagnosis of various skin tumors

The criteria for the diagnosis of various tumors were as described earlier.20 Papillomas were identified by their cauliflower-like exophytic tumorous growth with a narrow or broad base consisting of a series of connective tissue folds covered by stratified squamous epithelium, usually without cellular atypia. Squamous cell carcinomas are composed of very irregular endophytic epithelial growth with frequent cellular atypia. They are usually very differentiated, with the formation of keratinized layers of horny pearls.

Experimental protocol

For biochemical studies, two groups of mice (6 mice in each group) were taken. Group I received a single intramuscular injection of saline whereas group II received an intramuscular injection of imferon (5 mg iron/0.2 mL saline/mouse). 24 hours after the injection of saline/imferon all the animals were killed by cervical dislocation. Their dorsal skin was excised and processed for tissue preparation.

Tissue preparation for biochemical assays

After the desired time period, control and treated animals were sacrificed by cervical dislocation. The animals were immediately dissected to remove their skins which were washed in ice cold saline (0.85% NaCl). The subcutaneous layer of skin was completely scraped off with a scalpel blade to separate the epidermal layer by keeping the skin sample upside down on a petri dish containing crushed ice. The washed tissue was blotted between the folds of a filter paper and weighed on a metler balance. For biochemical studies a known amount of tissue was taken, minced properly and homogenized in appropriate buffer keeping the samples inside the crushed ice. Tissues were homogenized in a polytron homogenizer (Kinematica A.G.).

Estimation of tissue iron level

For estimating tissue iron levels in normal and iron overload states, blood samples were taken...
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overloaded animals, the animals were divided in two
groups of six mice each. Group I animals received a daily
intramuscular injection of saline for fourteen days whereas
group II animals received an intramuscular injection of
imferon (1 mg Fe/mouse) daily for a period of fourteen
days. 24 hours after the last injection of saline or imferon,
all animals were killed and their tissues removed and
processed for iron estimations. Just before sacrificing,
blood was withdrawn from all the animals and kept in
separate tubes for the separation of serum in order to
estimate iron levels.

Assay of lipid peroxidation

The assay for microsomal lipid peroxidation was done
according to the method of Wright et al.21 The reaction
mixture in a total volume of 1.0 mL contained 0.58 mL
phosphate buffer (0.1 M, pH 7.4), 0.2 mL microsome
obtained from the homogenate (10% w/v), 0.2 mL ascorbic
acid (100 mM), and 0.02 mL ferric chloride (100 mM).
The reaction mixture was incubated at 37°C in a shaking
water bath for one hour. The reaction was stopped by the
addition of 1 mL of TCA (10%). Following addition of 1.0
mL TBA (0.67%), all the tubes were placed in a boiling
water bath for a period of twenty minutes. In the end, tubes
were shifted to crushed ice bath and then centrifuged at
2500 g for 10 minutes. The amount of malonaldehyde
formed in each of the samples was assessed by measuring
the optical density of the supernatant at 535 nm using a
spectrophotometer (Milton Roy-21D) against a reagent
blank. The results were expressed as nmol MDA formed/
hr/gm of tissue at 3rC by using a molar extinction
coefficient of 1.56 x 10^5 M^-1 cm^-1.

Catalase activity

Catalase activity was assayed by the method of
Claiborne.22 Briefly, the assay mixture consisted of 1.95
mL phosphate buffer (0.05 M, pH 7.0), 1.0 mL hydrogen
peroxide (0.019 M), and 0.05 mL PMS (10% w/v) in a
total volume of 3.0 mL. Changes in absorbance were
recorded at 240 nm. Catalase activity was calculated in
terms of nmol H_2O_2 consumed/min/mg of protein.

Xanthine oxidase (XOD)

XOD catalyzes the conversion of xanthine to uric acid
which has a characteristic absorption peak around 290 nm.
The following spectrophotometric method based on the
modified procedure as described by Striee and Corta23 is
used here. The reaction mixture containing 0.2 mL PMS
diluted to 1 mL with buffer was incubated for 5 minutes at
37°C. Reaction was started by adding 0.1 mL xanthine.
The reaction mixture was kept at 37°C for 20 minutes.
The reaction was terminated by the addition of 0.5 mL ice-
cold perchloric acid (40%). After ten minutes, 2.5 mL
distilled water was added to the precipitated mixture

RESULTS

The effect of iron overload in various organs and
serum of mice is shown in Figure 1. Iron overload produces
a slight increase in the serum iron whereas tissue iron
levels show a marked increase as compared to saline-
treated control animals. In skin, iron levels show a nearly
three-fold increase whereas, in liver, it was more than
four-fold.

The effect of iron overload in mice skin-induced lipid
peroxidation, xanthine oxidase and catalase activities in
normal and iron overloaded mice skin is given in Figure 2.
The iron overloaded animals augmented lipid peroxidation
in comparison to saline alone treated animals. Increased
xanthine oxidase activity was observed in iron overloaded
animals, while catalase activity shows a decrease in iron
overloaded animals up to a level of less than 60% as
compared to the saline-treated control group.

The effect of iron overload on DMBA-induced
DMBA is activated by specific enzyme reactions to exert its neoplastic effects. It is metabolized by sequential reactions catalyzed by the cytochrome P-450 IA1-dependent monooxygenase system and epoxide hydrolase to its diol derivative, a compound which can be further metabolized by the two distinct pathways of epoxidation. The stereochemistry of epoxidation by the cytochrome P-450 dependent monooxygenase system and by free radicals (peroxyl radicals) is distinct and produces unique diol epoxides. Interestingly, both of these pathways exist in the mammalian skin. The DNA lesions produced as a result of either enzyme-dependent or free radical-dependent mechanisms are crucial for tumor development. Many of these metabolites may be converted into toxic quinones which oxidatively transfer an electron to produce superoxide anions. Thus, besides being initiators of carcinogenesis, they also produce tumors by exerting a promoting effect. In the present study, the observed decrease in the latency period of tumor induction indicates that iron overload exerts a promoting effect on the DNA lesions produced by DMBA. Since iron enhances skin lipid peroxidation and generates an oxidative stress in the skin, the higher production of free radicals may be responsible for the observed enhancement in the number and incidence of tumors in iron overloaded animals as compared to animals having normal iron levels. In addition, the observed increase in LPO and XOD activities in iron overloaded animals along with the decreased CAT activity in cutaneous tissues are suggestive of the tumor promoting effect of iron overload. The effect of iron overload on the tumor promoting response is evident from the data of the present study where enhanced activity of xanthine oxidase along with lipid peroxidation and diminished catalase activity was observed in iron overloaded animals. This is also supported by the fact that in the present study, increased iron overload-mediated enhancement of oxidant enzyme activities and enhanced iron overload-mediated decrease in the antioxidant enzyme activities were seen. It therefore seems that iron overload provides similar promoting potential as can be observed with a number of oxidants. This may be because of the production of in situ peroxidants by the iron overload in the tissue. Our results of the high cutaneous iron levels also corroborate the tumor data further suggesting an involvement of the iron-mediated biochemical cascade in the development and enhancement of skin tumors. In our previous studies, we have shown that in situ generated reactive oxygen species as a result of skin porphyrin photosensitization acts as a mild tumor promoting agent. The present data also support our findings that oxidants generated by iron overload may be acting as a modest tumor promoter, thus enhancing the tumor yield in iron overloaded animals. Our data, therefore, show that iron overload enhances DMBA-induced carcinogenesis in skin and suggest that oxidative stress generated by iron overload may be responsible for the observed augmentation of cutaneous tumorigenesis.

REFERENCES

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