

# LOCAL PRODUCTION OF PRIMARY POWDER OF *BACILLUS THURINGIENSIS* SEROTYPE H-14 IN IRAN AND DETERMINATION OF ITS INSECTICIDAL PROPERTIES AGAINST *CULEX PIPIENS* AND *ANOPHELES STEPHENSI*

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## ABSTRACT

Three media formulated from molasses and cornsteep liquor and another media based on the basal medium formulation of IPS-82 were assessed for the growth and production of insecticidal properties of *B. thuringiensis* H-14. Bacterial powders prepared from the broth cultures were assayed against the larvae of *Culex pipiens* and *Anopheles stephensi*. A standard powder of IPS-82 was included in the assay for comparison. Good growth was obtained in all the media and all powders were effective against the two types of mosquito larvae. Media with the basal medium formulation of Institute Pasteur Paris were the most effective. Molasses III media was also very favorable. The concentrations required to kill 50% of the larvae of *Culex pipiens* and *Anopheles stephensi* for these two were as follows:

- Media with the basal medium formulation of Institute Pasteur Paris:  
24hr. Lc50 and its 95% confidence limits of *Culex pipiens* and *Anopheles stephensi* = 0.0025, 0.0031, 0.0018 mg/lit  
0.1792, 0.2603, 0.1508 mg/lit
- Molasses III Media:  
24hr. Lc50 and its 95% confidence limits of *Culex pipiens* and *Anopheles stephensi* = 0.0103, 0.0115, 0.0092 mg/lit  
0.1994, 0.2223, 0.1769 mg/lit

The investigation shows that these media can be used for the production of *B. thuringiensis* H-14 primary powder.

*MJIRI, Vol.2, No.3, 229-236, 1988*

## INTRODUCTION

Malaria still takes a heavy toll on human life and suffering. Soon after the second world war the WHO recognised that malaria not only killed more people than any other disease but also interfered with the

development of agriculture and industry, especially in the new independent countries. However, during the past decade there has been a considerable increase of malaria in several tropical areas.<sup>1</sup> Malaria is still prevalent in the south eastern parts of Iran. In recent years, chloroquine-resistant strains of *Plasmodium falcipar-*

*um* in south east Asia have continued to spread westward and have been officially reported up to the southern border of Iran.<sup>2</sup> Some of the resistant cases in Iran were observed among Afghan immigrants or Pakistani individuals or Iranian Baluch tribes.<sup>3</sup> Establishment of the resistant strain in the south eastern parts of Iran can play an important role in spreading the infection between the uninfected vectors.

Mosquito vectors of diseases such as malaria are increasingly difficult to control, due in particular to the development of resistance to chemical insecticides.<sup>4</sup> It has become increasingly evident that control of disease vectors can't be based solely on chemical insecticides in view of the problem presented by the insects' resistance to insecticides, rising cost which limits adequate application in many countries, polluting the environment and destroying both harmful and harmless insects.<sup>5,6</sup> As mentioned, resistance to antimalarial drugs and the resistance of vectors to chemical insecticides are the important problems which indicate the importance of vector control.

Because of the above said reasons, scientists turned their attention to biological control mechanisms. Today biological control is regarded as a desirable technique for controlling insects, due to its minimal environmental impact and preventing the development of resistance in vectors. The entomopathogenic microorganism *Bacillus thuringiensis* is of the most promising biological control agent of pest and insect management since many strains are toxic specifically for lepidopterans and strain *Bacillus thuringiensis* serotype H-14 is highly toxic to dipterans. The first data on the existence and activity of the new strain *B. thuringiensis* H-14, appeared in 1977.<sup>7</sup>

*B. thuringiensis* H-14 is a spore-forming bacterium that produces one or more crystals of toxic protein (delta endotoxin) with each spore. The entomocidal properties of this protein crystal of *B. thuringiensis* H-14 have been shown to hold great promise as highly selective for mosquito and simuliid vectors of major parasitic diseases such as malaria and filariasis.<sup>6</sup> In many countries in which disease borne by mosquitoes are endemic, local production of *B. thuringiensis* H-14 has an economic advantage.<sup>4</sup> *B. thuringiensis* H-14 is highly host specific and practically harmless to non-target fauna, including the natural enemies of vectors which survive and augment its impacts. *B. thuringiensis* H-14 is also very safe to man and other vertebrates.<sup>5</sup>

In this experiment therefore, three media with different formulations based predominantly on locally available substrates in Iran such as molasses and corn-steep liquor were assessed as media for the local production of primary powders of *B. thuringiensis* H-14. Another media based on the basal medium formulation of Institute Pasteur Paris was used for the

production of a standard to be quite similar up to 90% to IPS-82 (International Pasteur Standard-82). These productions are then bioassayed against the larvae of *Culex pipiens* (one of the filariasis vectors) and *Anopheles stephensi* (malaria vector). This project is still continuing and the next results will be published in the future.

## MATERIALS AND METHODS

**Source of organism** Three flasks of standard powder IPS-82 were kindly granted by Professor H. DeBarjac, Institute Pasteur Paris. *B. thuringiensis* H-14 was isolated from this standard powder and was grown on brain heart infusion agar plate at 37°C for 24hr and stored at 4°C until used.

**Media** Three media designated Molasses I, II, III<sup>8</sup> and a medium based on basal medium formulation of IPS-82 were used for growth of *B. thuringiensis* H-14 in shake flask cultures. The pH of each medium was adjusted to 7-7.5 with %20 NaOH. Media were dispensed into 500ml Erlenmeyer flasks and sterilized at 121°C for 20 min (Table I).

**Seed Culture** Two loopfulls of *B. thuringiensis* H-14 from the BHIA plate was used to inoculate 50ml of TSB (trypticase soy broth) in 500ml flasks. TSB was used as a seed culture which was placed on the shaker (IRC-IU-Adult Kuhner AG, Co. Ltd.)

**Production flasks** Production flasks (500ml) containing 100ml of medium Molasses I, Molasses II, Molasses III each were inoculated with 100ml of the seed culture and incubated until 72 to 90% sporulation was obtained (after 72hr). Cultures were examined microscopically for sporulation at 24hr. intervals by staining spores with Schaeffer and Fulto's Method. Production flasks (500ml) containing 130ml of medium similar to standard 82 were inoculated with a full grown agar of the same agar media and was treated as Molasses media.

**Recovery** Four production media were recovered as bacterial primary powders at the end of fermentation by the freeze-drying technique (Lyovac GT3, Leyboldharius Ltd.) and were examined with electron microscope (E.M.10c, Zeiss) for the detection of crystals.

**Total viable cell count and spore counts** Total viable cell and spore counts were determined in the final whole culture by the pour plate method. Serial decimal dilutions of the final whole cultures were made in sterile %1 pepton water (oxid) and 0.5ml of each dilution in triplicate was added to a petridish, followed by the addition of 10ml of plate count agar (oxid) at 45°C. The culture and agar were mixed thoroughly and allowed to set. Plates were incubated at 32±2°C for 24 to 48hr, then counted with a colony counter. For spore

**Table I. Formulation of three molasses media<sup>(a)</sup> and media with the basal medium of Institute Pasteur Paris.**

Medium	Formulation
Molasses I	molasses: 1 <sup>ml</sup> cornsteep: 10 <sup>ml</sup> PO <sub>4</sub> H <sub>2</sub> K: 2 <sup>gr</sup> made up to 100 <sup>ml</sup> with D.W.
Molasses II	molasses: 0.5 <sup>ml</sup> cornsteep: 5 <sup>ml</sup> PO <sub>4</sub> H <sub>2</sub> K: 2 <sup>gr</sup> made up to 100 <sup>ml</sup> with D.W.
Molasses III	molasses: 0.5 <sup>ml</sup> SO <sub>4</sub> Mg. 7H <sub>2</sub> O: 1.23 <sup>gr</sup> SO <sub>4</sub> Mn. 1H <sub>2</sub> O: 0.016 <sup>gr</sup> SO <sub>4</sub> Zn. 7H <sub>2</sub> O: 0.14 <sup>gr</sup> Cl <sub>2</sub> Ca. 2H <sub>2</sub> O: 1.47 <sup>gr</sup> PO <sub>4</sub> H <sub>2</sub> K: : 3.4 <sup>gr</sup> Bactopecton: 0.7 <sup>gr</sup> made up to 100 <sup>ml</sup> with D.W.
Basal medium of Institute Pasteur Paris	
Solution 1	PO <sub>4</sub> H <sub>2</sub> K: 68 <sup>gr</sup> made up to 1000 <sup>ml</sup> with D.W.
Solution 2	SO <sub>4</sub> Mg. 7H <sub>2</sub> O: 12.3 <sup>gr</sup> SO <sub>4</sub> Mn. 4H <sub>2</sub> O: 0.223 <sup>gr</sup> SO <sub>4</sub> Zn. 7H <sub>2</sub> O: 1.4 <sup>gr</sup> made up to 1000 <sup>ml</sup> with D.W.
Solution 3	(SO <sub>4</sub> ) <sub>3</sub> Fe <sub>2</sub> : 2 <sup>gr</sup> SO <sub>4</sub> H <sub>2</sub> N: 100 <sup>ml</sup> made up to 1000 <sup>ml</sup> with D.W.
Solution 4	Cl <sub>2</sub> Ca. 4H <sub>2</sub> O: 18.3 <sup>gr</sup> with D.W. made up tp 1000 <sup>ml</sup> with D.W.
Preparation of the Basal medium	100ml of Solution 1 + 10 ml of Solution 2,3,4 7.5gr Bactopecton %30 glucose

(a) 100ml evaporated cornsteep was equal to 8.5gr solid material.  
Total molasses carbohydrates were measured by «Somogy Micro Copper-Method» and was equal to %47.

**Table II. Degree of sporulation, pH, total viable cell count and spore count in final whole cultures of three Molasses media<sup>(a)</sup> & media<sup>(b)</sup> with the basal medium formulation of Institute Pasteur Paris.**

Medium	% Sporulation	pH	T.V.C.C. <sup>(c)</sup> /ml	S.C. <sup>(d)</sup> /ml
Molasses I	88-91	8-8.5	1.5×10 <sup>9</sup>	1.3×10 <sup>9</sup>
Molasses II	72-80	7-7.5	1×10 <sup>9</sup>	7.5×10 <sup>8</sup>
Molasses III	92-95	7-7.5	10×10 <sup>9</sup>	9.3×10 <sup>9</sup>
media with the basal medium formulation of Institute Pasteur Paris	97-100	7-7.5	36×10 <sup>15</sup>	35.3×10 <sup>15</sup>

a = after 72hr.  
b = after 48hr.  
c = T. V. C. C. (total viable cell count)  
d = S. C. (spore count)

counts, cultures were pasteurized at 65°C for 20min before serial dilutions were made (Table II).

*Biossay of B.thuringiensis H-14 primary powders against mosquito larvae* Primary powders of *B.thuringiensis* H-14 produced from the four media were assayed against laboratory-rared third instar larvae of *Culex pipiens* and *Anopheles stephensi*. A standard powder of IPS-82 was included in the assay for comparison. The bioassay principle technique is exactly based on the bioassay method for the titration of *B.thuringiensis* H-14preparations with IPS-82 standard. Plastic cups of 200<sup>cc</sup> were prepared with 150ml free chlorine water with 25L<sub>3</sub> larvae of *Culex pipiens* or *Anopheles stephensi* in each cup. 50mg of each powder was suspended in distilled water and exact dilution of the suspension

## Insecticidal Properties against *Culex* and *Anopheles*

**Table III. Larvicidal activities of *B.thuringiensis* H-14 powders produced from local materials and basal medium formulation of Pasteur Institute standard in contrast with IPS-82 against *Culex pipiens* larvae after 24hr.**

Powder from medium and conc. mg/lit	Average % mortality at 24hr.	L <sub>50</sub> mg/lit	95% confidence limit		χ <sup>2(a)</sup>
			upper	lower	
Molasses I					
0.001	0	0.195	0.0215	0.0179	4.090
0.003	1				
0.005	5				
0.008	10.7				
0.01	15.7				
0.02	57.3				
0.03	73				
0.04	80.3				
Molasses II					
0.001	0	0.0171	0.0188	0.0155	9.641
0.003	2.7				
0.005	13				
0.008	16				
0.01	19				
0.02	65.7				
0.03	74				
0.04	83.7				
Molasses III					
0.001	0	0.0103	0.0115	0.0092	27.611
0.003	5				
0.005	23				
0.008	47.3				
0.01	66.3				
0.02	75				
0.03	79.3				
0.04	84.3				
Media with the basal medium formulation of Institute Pasteur Paris					
0.001	37	0.0025	0.0031	0.0018	3.524
0.003	48				
0.005	61.3				
0.008	72.7				
0.01	79.7				
0.02	86.3				
0.03	91				
0.04	92				
IPS-82					
0.001	41	0.0017	0.0021	0.0013	22.561
0.003	61.7				
0.005	74.7				
0.008	81				
0.01	98.3				
0.02	99.7				
0.03	100				
0.04	100				

(a): df = 6

\* critical value = 12.6

were added to the cups in order to obtain final concentrations of 0.04, 0.03, 0.02, 0.01, 0.008, 0.003, 0.001 mg/lit powders for *Culex pipiens* and concentrations of 0.4, 0.3, 0.2, 0.1 mg/lit of powders for *Anopheles stephensi* tests. 4 cups were used per dilution. Controls

consisted of 4 cups each containing 150ml D.W. and 25 larvae for each powder assayed. No food was added. Each experiment was incubated at 25±2°C and each assay was repeated three times. Mortality counts were made at 24 and 48hr.

**Table VI. Larvicidal activities of *B.thuringiensis* H-14 powders produced from local materials and basal medium formulation of Pasteur Institute standard in contrast with IPS-82 against *Anopheles stephensi* larvae after 48hr.**

powder from medium and conc. mg/lit	Average % mortality at 24hr	L <sub>c</sub> 50 mg/lit	95% confidence limit		X <sup>2(a)</sup>
			upper	lower	
Molasses I					
0.001	0	0.0105	0.0114	0.0097	4.533
0.003	2				
0.005	19				
0.008	33.3				
0.01	41.3				
0.02	85				
0.03	94.3				
0.04	98.7				
Molasses II					
0.001	0	0.0091	0.0099	0.0083	3.146
0.003	4				
0.005	23				
0.008	44.3				
0.01	56.7				
0.02	86.7				
0.03	93				
0.04	99				
Molasses III					
0.001	0	0.0056	0.0061	0.0051	38.726*
0.003	6.7				
0.005	56.3				
0.008	78				
0.01	86				
0.02	95.3				
0.03	97.7				
0.04	99				
Media with the basal medium formulaton of Institute Pasteur Paris					
0.001	40.3	0.0020	0.0025	0.0016	14.077
0.003	50.7				
0.005	67.7				
0.008	76				
0.01	91.7				
0.02	95				
0.03	96.3				
0.04	97				
IPS-82					
0.001	51.7	0.0012	0.0015	0.0008	23.537
0.003	70.3				
0.005	78.7				
0.008	85				
0.01	99.7				
0.02	100				
0.03	100				
0.04	100				

(a): df = 6

\* critical value = 12.6

*Calculation of results* The average percentage of mortality at 24 and 48hr for each dilution of each powder was converted to probits with the probit transformation table and the regression line relating probits

to log dose was calculated with a desk-calculator (Sx-30 cannon canula) to obtain L<sub>c</sub>50 results and its confidence limits. Mortality among control larvae was very rare and not more than 4% (Table III,IV,V,VI).

**Table V. Larvicidal activities of *B.thuringiensis* H-14 powders produced from local materials and basal medium formulation of Pasteur Institute standard in contrast with IPS-82 against *Anopheles stephensi* larvae after 24hr.**

Powder from medium and conc. mg/lit	Average % mortality at 24hr	Lc50 mg/lit	%95 confidence limit		$\chi^2_{(a)}$
			upper	lower	
Molasses I					
0.1	0	0.5637	0.8712	0.4659	1.775
0.2	5.6				
0.3	11.3				
0.4	30.6				
Molasses II					
0.1	1.3	1.834	62.37	1.1721	0.528
0.2	2.3				
0.3	6.3				
0.4	10.3				
Molasses III					
0.1	20.6	0.1994	0.2223	0.1769	1.369
0.2	51				
0.3	65.3				
0.4	83.3				
Media with the basal medium formulation of Institute Pasteur Paris					
0.1	33.3	0.1792	0.2063	0.1508	3.361
0.5	48				
0.3	66.6				
0.4	81.3				
IPS-82 <sup>(b)</sup>					
0.001	45	0.0016	0.0019	0.0012	24.128
0.003	52				
0.005	79				
0.008	95				
0.01	97				
0.02	98				
0.03	99				
0.04	100				

(a): df = 2  
critical value = 6

(b): df = 6  
\* critical value = 12.6

## RESULTS AND DISCUSSION

*B.thuringiensis* H-14 produced an appreciable amount of spores and delta endotoxin crystals in the four media between 48 and 72hr of fermentations. The degree of sporulation ranged from 72% in medium Molasses II to 95% in medium Molasses III. The T.V.C.C. were varied from  $1 \times 10^9$  in Molasses II media to  $10 \times 10^9$  in Molasses III media, and T.S.C. varied from  $7.5 \times 10^8$  in Molasses II to  $9.3 \times 10^9$  in Molasses III media. T.V.C.C. of 48hr medium with the basal medium formulation of Institute Pasteur was  $36 \times 10^{15}$ /ml with the degree of 97-100% sporulation after 48hr. Produced powders of *B.thuringiensis* H-14 were more effective against *Culex pipiens* than *Anopheles stephensi*;

similar observations have been reported by various workers.<sup>9-12</sup> The bioassay results at 24 and 48hr are summarized in Tables III for the two mosquito species. Maximum numbers of dead mosquito larvae were recorded in all *B.thuringiensis* H-14 powders at 48hr. Increase in larvae death after this period was very small. The insecticidal activities of the *B.thuringiensis* H-14 powders produced from local media did not correlate to the degree of sporulation observed in the final whole culture, and this has also been previously reported.<sup>13</sup>

The most effective locally produced *B.thuringiensis* H-14 primary powder was from medium Molasses III with the Lc<sup>50</sup> of 0.0103 and 0.0056 mg/lit after 24 and 48hr for *Culex pipiens* and 0.1994 and 0.1939 mg/lit for

Table VI. Larvicidal activities of *B.thuringiensis* H-14 powders produced from local materials and basal medium formulation of Pasteur Institute standard in contrast with IPS-82 against *Anopheles stephensi* larvae after 48hr.

Powder from medium and concn. mg/lit	Average %mortality at 24hr	Lc50 mg/lit	95% confidence limit		$\chi^2_{(a)}$
			upper	lower	
Molasses I					
0.1	4	0.3474	0.4419	0.3327	0.457
0.2	20.6				
0.3	36				
0.4	55.6				
Molasses II					
0.1	2	0.6301	0.9917	0.4968	0.667
0.2	7				
0.3	19.6				
0.4	30				
Molasses III					
0.1	23.3	0.1754	0.1939	0.1561	1.607
0.2	56.3				
0.3	74.3				
0.4	90				
Media with the basal medium formulation of Institute Pasteur Paris					
0.1	34.6	0.1561	0.1790	0.1306	1.420
0.2	58				
0.3	72.3				
0.4	86.3				
IPS-82					
0.001	45	0.0014	0.0018	0.009	6.816
0.003	58				
0.005	84				
0.008	100				
0.01	100				
0.02	100				
0.03	100				
0.04	100				

(a): df = 2  
critical value = 6

*Anopheles stephensi*. Primary powder of medium with the basal formulation of Institute Pasteur can be compared favorably with IPS-82 ( $Lc^{50}$  of 24 and 48hr IPS-82 for *Culex* and *Anopheles* were 0.0017, 0.0012, 0.0016, 0.0014 and  $Lc^{50}$  of 24 and 48hr primary powder of medium with basal formulation of Institute Pasteur were 0.0025, 0.0020, 0.1792, 0.1561 mg/lit).

Although the three molasses media are different only in amount of molasses and cornsteep, the extent to which they encouraged production of delta endotoxin crystal and its effectiveness against the two types of larvae was marked and quite favorable. This experiment shows that local or byproduct materials such as molasses and cornsteep in Iran or other countries can be used to produce *B.thuringiensis* H-14 powders with good activities against important mosquito species, especially vectors of malaria in endemic areas.

## REFERENCES

- 1- Chwatt-Leonard J B: Essential Malariology. Second Ed. Alden Press, Oxford, 1985.
- 2- WHO World Malaria Situation, World Health Statistics Quarterly 39: 171-205, 1986a.
- 3- Edrissian G H, Afshar A, Kanani A, Satvat M T, Ghorbani M: Resistance of *Plasmodium falciparum* to chloroquine in south eastern Iran. Medical Journal of the Islamic Republic of Iran. 1: 46-49, 1987.
- 4- Hertlein B, Hornby J, Levy R, Miller T Jr: Prospects of sporeforming bacteria for vector control with special emphasis on their local production potential. Mimeographed document WHO/VBC/80: 791;1980.
- 5- Rishikeh N, Burges H.D. Vandekar M: Operational use of *Bacillus thuringiensis* serotype H-14 and environmental safety. WHO/VBC/83: 871; 1983.
- 6- Margalit J, Dean D: The story of *Bacillus thuringiensis*. J Am Mosq control Assoc I(1): 4-7; 1985.
- 7- Carlberg G: *Bacillus Thuringiensis* and microbial control of flies.

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- Mircen Journal of Applied Microbiology and Biotechnology. II(1): 267-74; 1996.
- 8- Fayaz F, Moazami N: Three different media formulated from molasses and cornsteep liquor for the production of *Bacillus thuringiensis* serotype H-14. The Journal of the Pasteur Institute of Iran. 3: 54-60; 1987.
  - 9- Goldberg J, Margalit J: A bacterial spore demonstrating rapid larvicidal activity against *Anophele sergentii*, *Uranotaenia unguiculata*, *Culex univittatus*, *Aedes aegypti* and *Culex pipiens*, Mosquito News, 37: 3; 1977.
  - 10- De Barjac H: Une nouvelle Variete de *Bacillud thuringiensis* tres toxique pour les mostiques. *B.thuringiensis* var israelensis serotype 14. C.R. A:797-800; 1978.
  - 11- Garcia R, Desroches B: Toxicity of *Bacillus thuringiensis* var israelensis to some California mosquitoes under different conditions. Mosq. News 39:541-544; 1979.
  - 12- Tyrell D J, Davidson L I, Bulla L A, Ramsoda W A: A Toxicity of parasporal crystals of *Bacillus thuringiensis* subsp israelensis to mosquitoes. *Appl Environ Microbiol* 38:656-658; 1979.
  - 13- Dulmage H T. Production of spore delta endotoxin complex by variants of *Bacillus thuringiensis* in two fermentation media. *J Invertebr Pathol* 16:385-389.