Antibody response to glycan antigens of hydatid cyst fluid, laminated layer and protoscolex of Echinococcus granulosus

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Abstract

Background: Hydatid disease is characterized by long-term growth of hydatid cysts in the human. The glycan antigens have an important role in the immunological reaction of host sera to different glycan antigens of the cyst, has been investigated.

Methods: The antibody responses were tested to glycoprotein and glycolipid of the laminated layer (LL), cyst fluid (CF) and protoscolex (PS) antigens of E. Granulosus using ELISA and western immunoblotting tests. Thin-layer chromatography and β-elimination were used for glycan purification.

Results: Both hydatid cyst and normal human sera reacted with hydatid cyst fluid, protoscolices, laminated layer, glycoprotein and glycolipid antigens. The most antigen-antibody reaction was related to CF and PS antigens, and LL antigens had the minimal reaction with the sera. Thin layer chromatography (TLC) of the antigens showed presence of many glycan bands in the laminated layer.

Conclusion: The parasite may elaborate different glycan antigens in LL to evade host immune response.

Keywords: Echinococcus granulosus, Hydatid cyst, Laminated Layer, Thin-layer chromatography, Enzyme Linked Immunosorbent Assay, β-elimination

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Introduction

Hydatid disease or hydatidosis is a zoonotic disease caused by Larval stage of Echinococcus granulosus which is characterised by long-term growth of the cysts in the intermediate host (herbivores) (1-4). It has distributed on many continents, with the highest prevalence in Mediterranean areas and up to 75% of infected people may remain without symptom for more than 10 years (1, 2, 5-7). Hydatid cyst usually is located in the liver and/or the lung (8-10) and is filled with cyst fluid and protoscolices, the cyst wall which is composed of two layers: germinal layer and laminated layer (11). Each of these sectors (laminated layer, protoscolices and cyst fluid) has their own specific antigens that can be immunogenic or non-immunogenic (12, 13).

A hallmark of Echinococcus larva is their ability to survive within their hosts for a long time (14, 15). Despite host immune response, the parasite tries to escape the immune response and also down regulate the host defenses with different strategies such as antigenic mimicry, antigenic depletion and antigenic variation and it seems the immune response regulation in this helminth is beneficial for both the human host and the parasites (15-18). Parasite glycans have an important role in this regulation with “glycan gimmickry” strategy (19, 20). The unusual structure and the host-like glycan antigens of helmhths are the other mechanisms to escape from host immune system (19). Helminths produce different glycoproteins and lipids, and

↑What is “already known” in this topic:
Hydatidosis, as a zoonotic disease, has distributed on many continents with the highest prevalence in Mediterranean areas. Echinococcus multilocularis has a carbohydrate-rich laminated layer with a crucial role in the establishment of the infection in the mammalian host and protecting the parasite from immunological host reactions.

→What this article adds:
Glycan antigens of hydatid cyst have key roles in host-parasite relationship especially evasion from host immune system.
it has been shown that these compounds are used to regulate the host immune system (21, 22). For instance, *Echinococcus multilocularis* has a carbohydrate-rich laminated layer, which has a crucial role in the establishment of the infection in the mammalian host and protecting the parasite from immunological host reactions (23). In this context, it has been shown that in hydatid cyst infection some carbohydrate determinants are specific and immunogenic (24-26) and can immunoregulate the course of infection (27). Therefore, hydatid cyst glycan has been subjects of many research works recently. In this study immunological reaction of host sera with different glycan antigens of hydatid cyst has been investigated.

**Ethics statement**

This study was approved by the ethics committee of Isfahan Medical University, and sample collection was obtained with a written informed consent of patients and healthy controls.

**Methods**

In this descriptive research, the study population consisted of sera of either patient with hydatidosis or normal human sera. Normal sera (n=20) and hydatidosis sera (n=20) were collected from different hospitals in Isfahan, Iran. To prepare the antigens, liver and lung hydatid cyst of sheep were collected from Khomeini-Shahr slaughter house in Isfahan. At first, the hydatid cyst fluid was aspirated with a syringe and checked under the microscope for the presence of protoscolices. Following observation of the protoscolices, the cyst was included in the study. Aspirated cyst fluids were centrifuged for sedimentation of protoscolices. The supernatant was stored at -20 °C as hydatid cyst fluid, protoscolices, laminated layer, and germinal layers were collected from different hospitals in Isfahan, Iran. To prepare the antigens, liver and lung hydatid cyst of sheep were collected from Khomeini-Shahr slaughter house in Isfahan. At first, the hydatid cyst fluid was aspirated with a syringe and checked under the microscope for the presence of protoscolices. Following observation of the protoscolices, the cyst was included in the study. Aspirated cyst fluids were centrifuged for sedimentation of protoscolices. The supernatant was stored at -20 °C as hydatid cyst fluid, protoscolices, laminated layer, and germinal layers were collected from different hospitals in Isfahan, Iran. To prepare the antigens, liver and lung hydatid cyst of sheep were collected from Khomeini-Shahr slaughter house in Isfahan. At first, the hydatid cyst fluid was aspirated with a syringe and checked under the microscope for the presence of protoscolices. Following observation of the protoscolices, the cyst was included in the study. Aspirated cyst fluids were centrifuged for sedimentation of protoscolices. The supernatant was stored at -20 °C as hydatid cyst fluid, protoscolices, laminated layer, and germinal layers were collected from different hospitals in Isfahan, Iran. To prepare the antigens, liver and lung hydatid cyst of sheep were collected from Khomeini-Shahr slaughter house in Isfahan. At first, the hydatid cyst fluid was aspirated with a syringe and checked under the microscope for the presence of protoscolices. Following observation of the protoscolices, the cyst was included in the study. Aspirated cyst fluids were centrifuged for sedimentation of protoscolices. The supernatant was stored at -20 °C as hydatid cyst fluid, protoscolices, laminated layer, and germinal layers were collected from different hospitals in Isfahan, Iran. To prepare the antigens, liver and lung hydatid cyst of sheep were collected from Khomeini-Shahr slaughter house in Isfahan. At first, the hydatid cyst fluid was aspirated with a syringe and checked under the microscope for the presence of protoscolices. Following observation of the protoscolices, the cyst was included in the study. Aspirated cyst fluids were centrifuged for sedimentation of protoscl...
glycoprotein and glycolipid antigens. However, the mean OD of hydatid cyst patients’ sera was higher than that of normal sera. The highest difference between OD of hydatid cyst patients and normal sera related to glycoprotein antigens and the lowest difference related to glycolipid antigens. On the other hand, most antigen-antibody reaction was related to CF and PS antigens and LL antigens had minimal reaction with our sera (Table 1).

Various above antigens (LL, PS and CF) with different dilutions (1/2 and 1/4) run on 12% SDS-PAGE and stained with Coomassie blue (Figure 1). Figure 2 shows western immunoblotting of protoscolex (PS), laminated layer (LL) and cyst fluid (CF) probed with sera of patients with hydatid cyst or normal human serum. Laminin layer (LL), protoscolex (PS) and cyst fluid (CF) glycan antigens following β-elimination were subjected to TLC. Many glycan bands presented in the laminated layer (Figure 3).

Discussion
In our ELISA results, both sera of patients with hydatid cyst and normal human sera cross-reacted with different antigens of E. granulosus. These antigens may have an important role for the parasite to evade from the human immune system. Probably the parasite may elaborate these glycan antigens to raise antibodies that may block the specific sites for effective antibodies (32, 33). So, these antigens may have the potential to abolish production of specific immune responses. In this context, it has been shown that antibodies that Inhibit Malaria merozoite from invasion to erythrocyte are blocked by naturally acquired human antibodies and protozoa can attack to erythrocyte (34).

The laminate layer has a close contact with host tissues. However, according to our ELISA results antibody response to LL to hydatidosis was much lower than a response to protoscolices and cyst fluid. So it is possible that host antigens attach to the laminated layer surface. In agreement with this conception, it has been shown that E. granulosus have structural similarity to the host glycoproteins, which they effectively mimic like what we see in the other parasites to evade the immune system (19, 35).

High concentrations of glycol conjugate structure in the surface of hydatid cyst may also suppress several functions of the host immune system such as glycosylphosphatidylinositol and lipophosphoglycan as seen in protozoan parasites (36, 37). In TLC test, the strip of the LL Ag bands may indicate the presence of a wide range of different sugar in this layer, while PS Ag and CF Ag showed 2 bands or no band, respectively. In agreement with this result it has been shown that laminated layer is highly glycosylated (38-40).

The role of glycans in host immune evasion has also been shown in other helminths such as Schistosoma and filarial parasites (37). The tegument of S. mansoni contain an abundance of synthesize and adsorb host glycans molecules that use as a mechanism of immune evasion, and this helminth might disguise itself with the glycans against the attack of immune effectors (41). Filarial nematodes produce phosphorylcholine that anchors on the surface carbohydrate moieties and modulates host immune responses (42).

So in hydatid cyst, glycol conjugates may have a key function in host-parasite interaction such as protecting the parasite by regulating the host’s immune responses (19, 20, 38).
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Kanan JH and Chain BM, used the different protein bands of cyst fluid from SDS technique as a dendritic cells maturation and agents to regulate immune responses of the host (43). Also, it has been shown that immune evasion by hydatid cyst related to the production of different antibodies (44, 45). In the other hand, the carbohydrate-rich structure of LL of hydatid cyst is considered as protective material against host attack (46, 47). These observations are in agreement with our findings and confirm that glycan antigens of hydatid cyst have key roles in host-parasite relationship especially evasion from host immune system.

Conflict of Interests: None declared.

References