Introduction

Small fungal spores of the mold Aspergillus fumigatus (AF), which is abundant worldwide, can easily reach the lower airways and alveoli through inhalation (1, 2). In patients with asthma, the prevalence of Aspergillus sensitization is reported to be about 2% and it is more often associated with severe type of asthma (3). The major types of respiratory disease caused by these fungi are Aspergillus-induced asthma (AIA), severe asthma with fungal sensitization (SAFS), and allergic bronchopulmonary aspergillosis (ABPA) in patients with asthma (4, 5).

Patients with asthma who have an immediate positive reaction to the Aspergillus allergen without evidence of inflammation or tissue damage are considered to have AIA (2). The term SAFS was introduced in 2006 for patients with asthma who had an immediate positive skin reaction to Aspergillus antigens or elevated serum IgE levels to AF accompanied by frequent exacerbations of respiratory symptoms (4, 5).

ABPA is a complex pulmonary disease characterized by deterioration of lung function, elevated total serum IgE, elevated serum IgE and/or serum IgG to AF, recurrent transient chest X-ray infiltrate, and peripheral eosinophilia; nevertheless, the key sign is skin test reactivity to Aspergillus (6, 7). In patients with asthma, ABPA is sometimes diagnosed without evidence of typical proximal bronchiectasis; such cases are considered seropositive ABPA (ABPA-S) (6).

Shiraz is a city in Southwestern Iran which is influenced by dust storms originating from Arabic countries. The present study was designed to determine the frequency of sensitization to Aspergillus and the prevalence of ABPA and SAFS in individuals with severe asthma in Southwestern Iran.

This cross sectional study involved adult patients with severe asthma who were referred to the Allergy Clinic Center at Ali-Asghar hospital, affiliated to Shiraz University of
ABPA and SAFS in severe asthma

Med J Islam Repub Iran. 2019 (11 Sep); 33:95.

Medical Sciences in Shiraz, Iran, during a 6-month period from October 2014 to March 2015. All patients were diagnosed with severe asthma according to Expert Panel Report 3 criteria (8). After approval of the study protocol by the Ethics Committee of the university (EC-P-92-6337), written informed consent was obtained from each participant. Patients with a history of tuberculosis, smoking, diabetes mellitus, or hypertension and pregnant women were excluded from the study. Patients with asthma attack who received treatment with an oral glucocorticoid within the previous 6 months and patients with mild or moderate asthma were also excluded.

A questionnaire was used to obtain information about age, sex, duration of asthma, and the number of exacerbations in the past year, and each patient underwent physical examination.

Spirometry and bronchodilator reversibility tests were done for all patients. Skin prick tests were done for all patients with standard commercial extracts of AF (Greer, Lenoir, NC, USA). Histamine (10 mg/mL) and saline were used as positive and negative controls, respectively. Peripheral blood eosinophil count was done by standard hematoxylin and eosin staining, and an eosinophil count > 450/μL was considered to indicate eosinophilia.

Total serum IgE (Monobind, Lake Forest, CA, USA), serum specific IgE (Astra Biotech, Luckenwalde, Germany), and IgG (IBL International, Hamburg, Germany) to AF were measured with an enzyme-linked immunosorbent assay according to the manufacturer’s instructions. Total IgE > 200 IU/mL and specific IgE and specific IgG > 12 U/mL were considered positive.

A recent chest X-ray and high-resolution computed tomography (HRCT) scan of the chest were analyzed to identify bronchiectasis, parenchymal fibrosis, atelectasis, and mucus plugs.

Patients with asthma who had a positive immediate reaction to the Aspergillus allergen were classified into 3 diagnostic groups: (1) AIA; (2) SAFS: patients with severe asthma, ≥ 2 exacerbations of respiratory symptoms in the past year, and total serum IgE < 417 kU/L; and (3) ABPA. The minimum diagnostic criteria for ABPA in patients with asthma included deterioration of lung function, immediate cutaneous reactivity to AF, elevated total serum IgE > 417 kU/L or 1000 mg/mL, elevated serum IgE and/or IgG to AF, and chest X-ray infiltrate. Peripheral blood eosinophilia and central bronchiectasis on HRCT scan were considered additional diagnostic criteria for ABPA.

A total of 59 patients (39 women and 20 men, mean age 49±15 years, age range 17–79 years) with severe asthma were included in this study. The mean duration of asthma in these patients was 11±10 years. All patients had a history of cough, and physical examination disclosed bilateral wheezing and dyspnea at the time of the study. The number of patients with asthma who had ≥ 2 exacerbations of symptoms in the previous year was 50 (84.7%). All patients had airflow obstruction, and their FEV₁ ranged from 40% to 78%, and all showed reversibility in response to the bronchodilator.

The mean eosinophil count was 298± 243 /μL and blood eosinophilia (450/μL) was detected in 5 patients (8.4%). Increased total serum IgE levels were observed in 20 patients (34%) and 3 of them had a total IgE level > 417 kU/mL. Serum specific IgE and IgG against AF were positive in 15 patients (38.4%).

Infiltration was observed in chest X-rays in 15 patients (25.4%), and permanent findings in the form of parallel lines, ring shadows and consolidation were observed in 12 patients (20%). Chest HRCT revealed small-airway disease, bronchiectasis (3 patients with cylindrical and 1 patient with central proximal type), ground glass appearance in 4 patients, and pulmonary fibrosis in 2 patients. The prevalence of bronchiectasis in patients with asthma reportedly ranges from 18% to 52% (9).

The rate of sensitization to Aspergillus according to skin prick tests in the adult patients with severe asthma was seen in 10 patients (17%). An earlier study from the same geographical region reported positive skin reactions to fungi in 11% (25 out of 230) of the patients with childhood asthma (10). The difference in sensitization rates may be explained by the increased environmental exposure to indoor and outdoor fungal allergens during the lifetime of older patients. González-Díaz et al. reported that the rate of sensitization to fungi was 17.1% in 480 patients with respiratory allergy according to skin tests with fungal allergens (11). Mold allergies were found in 4.3% of 115 adult Turkish patients with asthma, with Cladosporium being the most common fungus (12). A study in Saudi Arabia on 139 patients with airway allergy found positive reactions to AF in 18.2% of the patients (13), a rate similar to that in patients in the present study. The prevalence of skin test reactivity to Aspergillus in patients with asthma has been reported to vary from 16% to 45% worldwide (14-16). This variation may be explained by differences in the AF extracts used for skin tests and differences in local humidity, which is related to fungal sensitization. The natural humidity in the region under study was below what was considered favorable to fungal growth. However, AF was found in household dust in 86% of homes in the study area, possibly because of the widespread use of air conditioners in homes (17). There was no significant relationship between age of patients and duration of asthma with a positive skin test.

The paraclinical data from these patients are summarized in Table 1. Six patients (10.2%) with a positive skin test to Aspergillus had ≥ 2 exacerbations of respiratory symptoms during the previous year and were diagnosed as having SAFS. Assuming that 10% of asthma cases are severe and a minimum rate of fungal sensitization of 33%, about 6.5 million people would be expected to have SAFS worldwide. However, the roles of regional or genetic factors in SAFS prevalence are still unknown (18).

Only 2 patients (3.4%) had all 4 main criteria and thus met the diagnostic criteria for ABPA in the present study. Other studies have reported a prevalence of ABPA, ranging from 1% to 27% among patients with asthma (6, 19, 20). For example, Al-Mobeireek et al. reported that the prevalence of ABPA was 2.7% among patients with asthma in Saudi Arabia (21). Although skin reactivity to Aspergillus in patients with asthma raises suspicions of ABPA, it is not solely confirmatory of this diagnosis. Large longitudinal
studies of patients with asthma and sensitization to AF may be needed to determine the progression of airway damage to better define SAFS and ABPA in the future.

According to the results, 3 patients had increased levels of total serum IgE > 417 kU/mL, whereas only 2 patients were considered to have ABPA based on the standard criteria. Elevated total serum IgE > 417 kU/mL is a major indicator in the diagnosis and management of ABPA. However, IgE levels may be increased in many patients with asthma but without ABPA because not all total serum IgE is directed to Aspergillus (6).

Among the patients, 2 out of 15 with high levels of Aspergillus-specific IgE were diagnosed as having ABPA and 5 were diagnosed as having SAFS. Aspergillus-specific IgE level can be considered the most sensitive test for ABPA in patients with asthma (22), but the diagnostic specificity of elevated Aspergillus-specific IgE levels in serum is limited for ABPA.

Of 10 patients with a positive Aspergillus skin test, 7 had increased levels of Aspergillus-specific IgE (kappa = 45%, P<0.001); the agreement between these 2 tests was lower than that reported by Agarwal et al (22). One recent study emphasized that both skin test results and specific IgE measurement should be considered in the diagnosis of fungal allergy because of discordance between the test results in approximately one fourth of patients with severe asthma (23). In this connection, Cohen’s kappa value showed strong concordance between positivity for serum specific IgE to Aspergillus and total IgE > 200 IU/mL (kappa = 73.2%, P = 0.013) in the present study.

Peripheral blood eosinophilia is present in patients with ABPA who are not taking oral corticosteroids at the time of exacerbation or during the acute phase of asthma. A peripheral blood eosinophil count > 1000 cells/μL is often considered a diagnostic criterion, although some authors consider it to be suggestive (rather than diagnostic) of ABPA (20). None of the patients had an eosinophil count > 1000 cells/μL. In the diagnostic criteria for ABPA recently proposed by the International Society for Human and Animal Mycology, a peripheral blood eosinophil count above 500 is considered a minor criterion which awaits prospective validation (24).

Bronchiectasis was found to be more prevalent among patients with ABPA than patients with asthma (25). One patient had central proximal bronchiectasis and another was diagnosed as having ABPA-S in this study. Results of this study were consistent with the following reports: Bronchiectasis was diagnosed in 4 patients but only 1 of them was diagnosed as having ABPA.

The prevalence of ABPA was low in this study. Considering the high prevalence of asthma and fungal sensitization in severe asthma in the area under study, physicians should be concerned about ABPA and SAFS in patients with severe asthma, especially those with a positive skin test to Aspergillus allergens.

### Acknowledgments

This work was supported by a grant from Shiraz University of Medical Sciences (Grant No. 92-6337). We thank K. Shashok (Author AID in the Eastern Mediterranean) for English editing of the manuscript.

### Conflict of Interests

The authors declare that they have no competing interests.

### References


http://mjiri.iums.ac.ir

Med J Islam Repub Iran. 2019 (11 Sep); 33.95.