ANTINOCICEPTIVE INTERACTION BETWEEN ADENOSINE AND CARBAMAZEPINE IN MICE

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ABSTRACT

There are several reports that adenosine and carbamazepine have pharmacodynamic interaction. In this study the antinociceptive interaction of these two agents was evaluated in mice by the hot plate test. Agents were injected intraperitoneally.

0.2 and 0.4 mg/kg doses of R-phenylisopropyladenosine (R-PIA) substantially showed antinociceptive effects. Carbamazepine had antinociceptive actions except at a dose of 0.8 mg/kg. Theophylline (12.5 mg/kg), as a non-selective antagonist of adenosine receptors, showed a nociceptive effect. Dipyridamole, an uptake blocker of adenosine, had no antinociceptive effect even with a dosage of 90 mg/kg. Theophylline did not decrease the antinociceptive effect of carbamazepine, while dipyridamole increased the antinociceptive effect of carbamazepine 30 minutes after administration. Carbamazepine (8 mg/kg) inhibited the antinociceptive effect of R-PIA.

This study supports the possibility that the interaction between adenosine and carbamazepine may be related to their actions at adenosine receptor sites in the brain. MJIRI, Vol. 12, No. 4, 381-386, 1999

Keywords: Adenosine, R-PIA, Carbamazepine, Theophylline, Dipyridamole, Antinociceptive.

INTRODUCTION

Adenosine has effects on almost all kinds of mammalian tissue including the central nervous system. As a neuromodulator adenosine has a profound depressant action in the central nervous system. Its action may be mediated pre- and post-synaptically through receptor-mediated mechanisms, including effects on second messenger systems, transmembrane ion fluxes and neurotransmitter release. Adenosine (5'-AMP) is the active form of adenosine that binds to G proteins to activate adenylate cyclase. Adenosine analogs like 2'-deoxyadenosine and 3',5'-cyclic adenosine mono-phosphate (cyclic AMP) are similar in structure to adenosine and are substrates for adenosine deaminase. Adenosine analogs may interfere with the deamination of adenosine and can therefore increase the concentration of adenosine within cells. The presence of adenosine receptors may increase the activity of adenosine in the brain. Adenosine receptors are located in the hippocampus, cerebral cortex, thalamus, hypothalamus, substantia nigra, dorsal horn of the spinal cord, and the retina. Adenosine receptors have been identified at these sites by autoradiography with radioligands such as [3H]N6-phenylisopropyladenosine (PIA). The PIA receptor is a G protein-coupled receptor that is coupled to adenylyl cyclase. The binding of adenosine or adenosine analogs to the PIA receptor leads to an increase in the intracellular cyclic AMP concentration. Antagonists of the PIA receptor such as theophylline and pentostatin prevent the increase in cyclic AMP concentration.

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The mechanism of the anticonvulsant activity of carbamazepine is not fully clear. This drug, like phenytoin, inhibits voltage-dependent Na+ channels. Interest in the possibility that the anticonvulsant action of carbamazepine may relate to an interaction with adenosine receptors developed as a result of different observations. Since the clinical properties of carbamazepine are reminiscent of the anticonvulsant, sedative, and anxiolytic properties of adenosine, an adenosine-agonistic activity of carbamazepine has been proposed. However, indirect evidence suggested a rather antagonistic activity of carbamazepine at adenosine receptors. Carbamazepine inhibits the binding of adenosine analogs [3H]N6-phenylisopropyladenosine and [3H]cyclohexyladenosine to synaptosomal membranes in the rat brain. Carbamazepine was found to antagonize the A1-receptor-
mediated inhibition of cyclic AMP accumulation in cultured astroblasts and in GH3-cells. Carbamazepine also inhibited the adenosine-induced increase in the level of cyclic AMP in cultured astroblasts, which is mediated by low-affinity A_2b receptors. In the present study we have investigated the antinociceptive interaction of carbamazepine with the adenosine receptor in the central nervous system.

MATERIAL AND METHODS

Animals
Male and female albino mice weighing 25-30g were obtained from a random bred colony maintained on a special diet (Khorassan Javane Co., Mashhad, I.R. Iran) in the animal lab of Mashhad University of Medical Sciences. Animals were housed in a room with a 12/12 hr light/dark cycle at 21±2°C. The animals had free access to food and water.

Measurement of analgesic activity
Antinociceptive activity was assessed by the hot-plate test. The temperature of a metal surface was maintained at 55±0.2°C. The latency to a discomfort reaction (licking paws or jumping) was determined before and after drug administration. The cut-off time was 40 sec. All drugs were injected intraperitoneally (ip). When two agents were used, injections in two sites of the peritoneum were with a 30 minute interval.

Carbamazepine was dissolved in 50% propylene glycol and 50% saline warmed on a water bath. The solvent of R-PIA was water. Theophylline was dissolved in NaOH and the final preparation was restored to a pH of 7.4. Dipyridamole was suspended in carboxymethyl-cellulose (1%). Control groups received only the solvent. Carbamazepine, dipyridamole and theophylline were gifts from CIBA-GEIGY, Thomae and Darupakhsh (I.R. Iran), respectively. R-PIA was purchased from Sigma.

Statistical analysis
The data were expressed as mean values±S.E.M. and tested with analysis of variance followed by the multiple comparison test of Tukey.

RESULTS

Effect of R-PIA on hot-plate latency
R-PIA significantly induced an antinociceptive effect and the maximum responses were observed at 0.4 mg/kg, 30 minutes after agent injection (Fig. 1).

Effect of carbamazepine on hot-plate latency
Intraperitoneal administration of different doses of carbamazepine to mice induced a dose-dependent
Table I. Effect of theophylline on carbamazepine (CBZ) antinociception in the hot-plate test. All mice were injected intraperitoneally.

<table>
<thead>
<tr>
<th>Treatment (mg/kg)</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>90 min</th>
<th>120 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (Theophylline)</td>
<td>6.28±0.39</td>
<td>7.12±0.36</td>
<td>6.13±0.45</td>
<td>6.75±0.30</td>
<td>7.05±0.55</td>
</tr>
<tr>
<td>Control (Carbamazepine)</td>
<td>4.17±0.23</td>
<td>4.47±0.38</td>
<td>4.17±0.23</td>
<td>4.90±0.36</td>
<td>5.22±0.17</td>
</tr>
<tr>
<td>Control (Theophylline+CBZ)</td>
<td>3.87±0.10</td>
<td>3.78±0.25</td>
<td>4.63±0.25</td>
<td>3.97±0.33</td>
<td>3.82±0.19</td>
</tr>
<tr>
<td>Theophylline (7)</td>
<td>5.19±0.38</td>
<td>4.84±0.35</td>
<td>4.27±0.23</td>
<td>3.93±0.58</td>
<td>4.57±0.56</td>
</tr>
<tr>
<td>Carbamazepine (20)</td>
<td>4.87±0.38</td>
<td>7.93±0.97</td>
<td>7.43±0.96</td>
<td>7.23±0.55</td>
<td>6.20±0.39</td>
</tr>
<tr>
<td>Carbamazepine (25)</td>
<td>4.08±0.21</td>
<td>8.30±0.55</td>
<td>9.46±0.93</td>
<td>8.45±1.02</td>
<td>7.45±0.53</td>
</tr>
<tr>
<td>Theophylline (7) + CBZ (20)</td>
<td>5.50±0.44</td>
<td>5.62±0.26</td>
<td>8.67±0.56</td>
<td>9.73±0.64</td>
<td>7.93±0.45</td>
</tr>
<tr>
<td>Theophylline (7) + CBZ (25)</td>
<td>5.64±0.22</td>
<td>5.76±0.17</td>
<td>11.63***±</td>
<td>10.49*±</td>
<td>8.59±0.48</td>
</tr>
</tbody>
</table>

*p<0.05, ***p<0.001; theophylline group compared with theophylline+CBZ 25 mg/kg group, Tukey-Kramer test, N= 6 mice; control group of carbamazepine and theophylline received propylene glycol+saline and sodium hydroxide, respectively.

antinociceptive effect (Fig. 2). The maximum response was observed with 25 mg/kg, 60 minutes after drug injection.

Effect of dipyridamole on hot-plate latency
Doses up to 90 mg/kg of dipyridamole had no effect on the pain threshold of mice (Fig. 3).

Effect of theophylline on hot-plate latency
Theophylline reduced hot plate latency, implying an enhancement of pain sensitivity. The maximum response was observed with 50 mg/kg, 30 minutes after drug injection (Fig. 4).

Effect of theophylline on carbamazepine-induced antinociception
Theophylline at 7 mg/kg increased the antinociceptive effect of carbamazepine (25 mg/kg) at 60 and 90 minutes after injection (Table I).

Effect of dipyridamole on carbamazepine-induced antinociception
Dipyridamole increased the antinociceptive effect of carbamazepine only at a dose of 90 mg/kg (Table II).

Effect of carbamazepine on R-PIA induced antinociception
Carbamazepine, at a concentration that had no antinociceptive activity (8 mg/kg), reduced the antinociceptive effect of R-PIA 30 and 60 minutes after its administration (Table III).

DISCUSSION

The results of this study indicate that carbamazepine inhibits the antinociceptive effect of R-PIA at a concentration that by itself has no effect.

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Table II. Effect of dipyridamole on carbamazepine (CBZ) antinociception in the hot-plate test. All mice were injected intraperitoneally.

<table>
<thead>
<tr>
<th>Treatment (mg/kg)</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>90 min</th>
<th>120 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (Dipyridamole)</td>
<td>7.08±0.39</td>
<td>5.60±0.48</td>
<td>5.83±0.45</td>
<td>5.80±0.46</td>
<td>4.58±0.24</td>
</tr>
<tr>
<td>Control (Carbamazepine)</td>
<td>4.17±0.23</td>
<td>4.42±0.43</td>
<td>4.17±0.28</td>
<td>4.90±0.39</td>
<td>5.22±0.24</td>
</tr>
<tr>
<td>Control (Dipyridamole+CBZ)</td>
<td>4.58±0.24</td>
<td>4.59±0.34</td>
<td>4.95±0.25</td>
<td>5.34±0.27</td>
<td>5.69±0.33</td>
</tr>
<tr>
<td>Carbamazepine (25)</td>
<td>4.08±0.21</td>
<td>8.30±0.55</td>
<td>9.46±0.99</td>
<td>8.45±1.02</td>
<td>7.45±0.53</td>
</tr>
<tr>
<td>Dipyridamole (60)</td>
<td>5.83±0.29</td>
<td>5.55±0.43</td>
<td>5.17±0.32</td>
<td>5.53±0.62</td>
<td>5.81±0.41</td>
</tr>
<tr>
<td>Dipyridamole (90)</td>
<td>4.27±0.28</td>
<td>5.02±0.69</td>
<td>5.18±0.33</td>
<td>5.82±0.42</td>
<td>5.32±0.65</td>
</tr>
<tr>
<td>Dipyridamole (60) + CBZ (25)</td>
<td>5.10±0.39</td>
<td>10.81±0.87</td>
<td>8.84±0.49</td>
<td>7.03±0.51</td>
<td>6.17±0.56</td>
</tr>
<tr>
<td>Dipyridamole (90) + CBZ (25)</td>
<td>5.98±0.32</td>
<td>11.58±0.74</td>
<td>10.18±0.43</td>
<td>8.90±0.42</td>
<td>8.90±0.42</td>
</tr>
</tbody>
</table>

***p<0.001; carbamazepine group compared with dipyridamole (90)+CBZ 25 mg/kg group, Tukey-Kramer test. N= 6 mice; control group of carbamazepine and dipyridamole received propylene glycol+saline and carboxymethyl cellulose, respectively.

The antinociceptive dose of carbamazepine in this study is similar to the dose which showed anticonvulsant activity against electroshock seizures in mice (18 mg/kg) or pentylenetetrazole-induced tonic extensor seizures in rats (10 mg/kg). Thus carbamazepine—anticonvulsant doses—may have antinociceptive effects.

Theophylline dose-dependently induced a reduction of hot-plate latency. This effect may relate to the blockade of A\textsubscript{1} adenosine receptors that have an analgesic effect. In this study theophylline increased the antinociceptive effect of carbamazepine. This interaction implies that the antinociceptive effect of carbamazepine was not mediated by adenosine receptors. The reason for this effect is not clear in this experiment. In the study of interaction of adenosine with the GABA system, theophylline increased the antinociceptive effect of baclofen. The authors of this article discussed that this effect may relate to changes in dopaminergic equilibrium.

In this study intraperitoneal injections of dipyridamole, an adenosine uptake blocker, had no antinociceptive action in mice. This may indicate that there is not enough adenosine in the extracellular medium in the normal situation. This neuromodulator is liberated during situations such as hypoxia or seizures when dipyridamole can then be effective. Dipyridamole increased the antinociceptive effect of carbamazepine only at 30 minutes after injection. The aim of this experiment was to study the role of adenosine in affecting the actions of carbamazepine. Since the effect of carbamazepine was increased only at 30 minutes, it is unlikely that adenosine has a role in the antinociceptive effect of carbamazepine. Carbamazepine blocks adenosine uptake and this may be potentiated by dipyridamole, but carbamazepine only has this effect with high doses.

At a concentration that had no antinociceptive effect, carbamazepine inhibited the antinociceptive effect of the A\textsubscript{1} adenosine receptor agonist, R-PIA. This interaction is...
similar to the results of Stone who showed that carbamazepine blocked the inhibitory activity of 2-chloroadenosine on orthodromic potentials of hippocampal slices.

In view of this fact that both adenosine and carbamazepine have depressant actions on the central nervous system, it is a paradox that carbamazepine blocks the antinociceptive effect of an adenosine agonist. However, other results such as the reduction of the anticonvulsant activity of adenosine on cortex neurons, inhibition of the effect of adenosine on adenylyl cyclase and reduction of adenosine analogue effects on muscle contraction by carbamazepine are consistent with our results. On the other hand, adenosine depresses the excitability of pyramidal neurons in the hippocampus. This effect was lost in calcium-free media and carbamazepine restored sensitivity to adenosine in calcium-free medium.

With chronic treatment, carbamazepine also increased adenosine A<sub>1</sub> receptor density in the rat brain. Therefore, chronic carbamazepine treatment appears to upregulate adenosine receptors, suggesting that this drug may act as an adenosine antagonist. In contrast to other anticonvulsants such as phenobarbital, phenytoin, primidone, sodium valproate and ethosuximide, the anticonvulsant carbamazepine inhibited binding of cyclohexyladenosine, a selective adenosine A<sub>1</sub> receptor, to rat cerebral tissues.

In summary, this study has demonstrated a reduction of adenosine antinociceptive effects by carbamazepine. Carbamazepine may antagonize adenosine A<sub>1</sub> receptors in the brain. Further basic and clinical studies are required to confirm this interaction.

**ACKNOWLEDGEMENT**

The authors are thankful to T.W. Stone, Professor of Pharmacology, Division of Neuroscience and Biomedical Systems, University of Glasgow, Scotland, for his guidance.

**REFERENCES**


### Table III. Effect of carbamazepine (CBZ) on R-phenylisopropyladenosine (R-PIA) antinociception in the hot-plate test. All mice were injected intraperitoneally.

<table>
<thead>
<tr>
<th>Treatment (mg/kg)</th>
<th>0 min</th>
<th>30 min</th>
<th>60 min</th>
<th>90 min</th>
<th>120 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (R-PIA)</td>
<td>7.87±0.56</td>
<td>7.57±0.61</td>
<td>8.23±0.55</td>
<td>8.04±0.58</td>
<td>8.33±0.51</td>
</tr>
<tr>
<td>Control (Carbamazepine)</td>
<td>8.33±0.67</td>
<td>7.25±0.43</td>
<td>8.10±0.61</td>
<td>8.33±0.48</td>
<td>7.10±0.42</td>
</tr>
<tr>
<td>Control (R-PIA+CBZ)</td>
<td>7.30±0.44</td>
<td>7.90±0.57</td>
<td>7.51±0.43</td>
<td>7.58±0.54</td>
<td>7.78±0.54</td>
</tr>
<tr>
<td>Carbamazepine (8)</td>
<td>6.97±0.50</td>
<td>7.77±0.55</td>
<td>9.16±0.42</td>
<td>9.49±0.47</td>
<td>8.31±0.34</td>
</tr>
<tr>
<td>R-PIA (0.4)</td>
<td>7.86±0.60</td>
<td>18.00±0.59</td>
<td>18.20±0.94</td>
<td>13.00±1.36</td>
<td>13.24±1.00</td>
</tr>
<tr>
<td>R-PIA (0.4) + Carbamazepine (8)</td>
<td>7.00±0.32</td>
<td>13.84***±</td>
<td>12.53±0.57</td>
<td>12.51±0.51</td>
<td></td>
</tr>
</tbody>
</table>

**p<0.01, ***p<0.001; R-PIA 0.4 mg/kg group compared with R-PIA+CBZ group, Tukey-Kramer test, N=6 mice; control group of R-PIA and carbamazepine received saline and propylene glycol+saline, respectively.
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