



## Current Insights into Plausible Mechanisms of Chromium (VI) Neurotoxicity in the Brain and Future Perspectives

Marat Izteuov<sup>1</sup>, Nazerke Abugaliyeva<sup>1\*</sup> , Sergey Ryzhkin<sup>2</sup>, Yerbolat Izteuov<sup>1</sup>, Samat Saparbaev<sup>3</sup>, Gaziza Smagulova<sup>1</sup>

Received: 16 Jul 2025

Published: 16 Dec 2025

### Abstract

**Background:** Hexavalent chromium (Cr (VI)) is a known neurotoxin and environmental contaminant. Despite its recognition, the underlying mechanisms by which Cr (VI) induces neurological damage remain insufficiently explored. The complexities of the Central Nervous System (CNS), including the Blood Brain Barrier (BBB) and supporting brain cells, contribute to regions-specific susceptibility within the brain. Understanding Cr (VI) neurotoxicity is crucial for its potential role in neurodegenerative diseases.

**Methods:** A Systematic Review was conducted using international databases (PubMed, Medline, Scopus, and Web of Science) and Google Scholar. Only open-access, free full-text articles published in English between 2010 and 2025 were included. Following PRISMA 2020 guidelines, a total of 19 relevant studies were selected, comprising 12 animal-based and 7 human cohort studies.

**Results:** Animal studies investigated the effects of Cr (VI) via various administration methods and doses, revealed evidence of oxidative stress, inflammatory markers, and apoptotic changes in the brain. Interventional studies showed delayed toxicity when antioxidant agents were used prior to Cr (VI) exposure, including PDC (Potassium Dichromate), SA (Sodium Alginate), and TNG (Tangeretin). Human studies, including autopsies and cell culture analyses, demonstrated neurotoxic effects in conditions such as ALS (Amyotrophic Lateral Sclerosis), nAMD (Neovascular Age-Related Macular Degeneration).

**Conclusion:** Animal studies have clarified the role of oxidative stress in Cr (VI)-induced neurotoxicity. Human cohort studies have identified Cr (VI) as an environmental risk factor for both neurodegenerative and neurobehavioral disorders. Future research should focus on defining harmful levels of Cr (VI) and exploring potential antioxidant therapies.

**Keywords:** Chromium, Toxicity, Heavy metals, Neurodegenerative disorders, Oxidative stress, Neuroinflammation

**Conflicts of Interest:** None declared

**Funding:** This study was conducted within the framework of the scientific project supported by grant funding from the Committee of Science of the Ministry of Science and Higher Education of the Republic of Kazakhstan, IRN AP2348980, titled "Prevention of Induced Radiation-Chemical (Chromium) Oncogenesis, Including Offspring in Experiment" (contract No. 308 GF 24-26, dated 09.09.2024, registration number 0124RK00949).

\*This work has been published under CC BY-NC-SA 4.0 license.

Copyright © [Iran University of Medical Sciences](#)

**Cite this article as:** Izteuov M, Abugaliyeva N, Ryzhkin S, Izteuov Y, Saparbaev S, Smagulova G. Current Insights into Plausible Mechanisms of Chromium (VI) Neurotoxicity in the Brain and Future Perspectives. *Med J Islam Repub Iran.* 2025 (16 Dec);39:157. <https://doi.org/10.47176/mjiri.39.157>

### Introduction

Chromium (Cr) is a heavy metal that naturally occurs in two primary oxidation states: trivalent chromium [Cr(III)] and hexavalent chromium [Cr(VI)]. Cr(III) is considered less harmful and may even be beneficial in trace amounts, whereas Cr(VI) is well known for its car-

cinogenic properties and is recognized as a major environmental contaminant (1-5). Industrial activities such as coal and oil combustion, the use of oxidizing pigments, paint manufacturing, fertilizer production, oil well drilling, metal plating, and leather processing con-

**Corresponding author:** Dr Nazerke Abugaliyeva, [nazerkeabugaliyevaaa@gmail.com](mailto:nazerkeabugaliyevaaa@gmail.com)

<sup>1</sup> West Kazakhstan Marat Ospanov Medical University, Aktobe, Kazakhstan

<sup>2</sup> Medical Academy of Continuous Professional Education, Kazan State Medical University, Moscow, Kazan, Russia

<sup>3</sup> MC Al-Jhami, Aktobe, Kazakhstan

#### ↑What is “already known” in this topic:

Hexavalent chromium (Cr (VI)) is widely recognized as a neurotoxin, that is associated with oxidative stress and neuronal damage. However, the mechanisms by which Cr (VI) induces neurotoxicity, particularly regarding its impact on the blood-brain barrier (BBB) and subsequent neurodegenerative diseases, remain underexplored.

#### →What this article adds:

This review consolidates the current understanding of Cr (VI) induced neurotoxicity, emphasizing experimental findings and human cohort studies. It highlights the role of Cr (VI) in oxidative stress, neuroinflammation, and neurodegenerative disorders, while also proposing future research directions and potential therapeutic approaches such as antioxidant interventions.

tribute substantially to environmental Cr(VI) pollution (6). Occupational exposure, primarily through inhalation, is strongly associated with an elevated risk of lung cancer, while ingestion of contaminated water has been linked to an increased risk of liver cancer. Once inside the body, Cr(VI) mimics sulfate and phosphate ions, allowing it to enter cells passively through ion channels (7). Within the cytoplasm, it is rapidly reduced to Cr(III), proceeding through intermediate oxidation states such as Cr(V) and Cr(IV) (8). This redox process generates reactive oxygen species (ROS), including superoxide anions, hydrogen peroxide, and hydroxyl radicals, all of which contribute to cellular oxidative stress. Furthermore, Cr(VI) has been shown to induce genomic instability by disrupting DNA repair pathways and damaging mitochondria both key mechanisms in carcinogenesis (9). Hexavalent chromium [Cr(VI)] is classified as a Group 1 carcinogen by the International Agency for Research on Cancer (IARC), confirming its carcinogenicity in humans (10). Despite this established toxicity, research into the neurological effects of Cr(VI) remains limited. The blood-brain barrier (BBB), which protects the brain from harmful substances, relies on the coordinated activity of astrocytes, microglia, pericytes, endothelial cells, and certain neurons, to maintain its structural integrity and selective permeability. This integrity is essential for preventing the entry of toxic agents such as heavy metals into the central nervous system (CNS) (11, 12). It remains unclear whether Cr(VI) can cross or damage the BBB. The detection of chromium in cerebrospinal fluid (CSF) from a hip transplant case in which it was identified as Cr(III) rather than Cr(VI)—suggests that chromium, in general, may have the potential to interact with or penetrate the CNS (13). An alternative route of entry into the brain could be through the olfactory bulb, which lies outside BBB protection (14).

Moreover, the hypothalamus and pituitary gland, which are involved in several homeostatic processes, maintain continuous contact with the blood stream and possess a weaker BBB (15). These regions could therefore represent potential entry points for Cr(VI). However, the mechanisms underlying Cr(VI)-induced neurotoxicity remain poorly understood. Although the toxic effects of Cr(VI) are well documented, there is limited occupational evidence linking chromium exposure to neurological or behavioral disorders (16). Animal studies have described the pathophysiological basis of chromium-induced neurotoxicity and highlighted the potential therapeutic relevance of certain agents (17–19).

Understanding these mechanisms could provide important insights for human studies investigating the association between Cr(VI) exposure and neurodegenerative disorders such as Alzheimer's or Parkinson's disease.

Therefore, this systematic review examines the existing literature on Cr(VI) toxicity and its adverse effects on the central and peripheral nervous systems. The specific objectives of this review are to:

1. Summarize experimental studies elucidating the possible mechanisms of nerve damage caused by Cr(VI) toxicity;
2. Summarize observational studies demonstrating the

clinical implications of Cr(VI) exposure;

3. And Identify research gaps and propose recommendations for future studies and guidelines.

## Methods

### Search Strategy

This systematic literature review was conducted in accordance with the PRISMA 2020 guidelines. A comprehensive search of relevant scientific literature was performed across major international databases, including PubMed, Scopus, Web of Science, and Embase, to ensure broad coverage of related studies. Additionally, Google Scholar was searched to capture freely available and gray literature. The search strategy combined Medical Subject Headings (MeSH) and free-text terms using Boolean operators (AND, OR), applying the following keywords: ("chromium" OR "Cr(VI)" OR "hexavalent chromium") AND ("brain" OR "central nervous system" OR "CNS") AND ("toxicity" OR "neurotoxicity" OR "oxidative stress" OR "inflammation" OR "apoptosis" OR "neurodegeneration"). The search covered studies published between January 2010 and March 2025, restricted to English-language, full-text, and open-access articles. To ensure completeness, the reference lists of relevant publications, as well as the "Cited By" and "Related Articles" sections of included studies, were also manually screened. A total of 14,264 records were initially identified across all databases. Of these, 13,902 were excluded through automated filtering and duplicate removal. After de-duplication, 38 articles were selected for eligibility assessment, of which 19 were retained for reporting and discussion. The remaining 18 articles were excluded for the following reasons: (1) Cr(VI) was not discussed; (2) neuronal tissues were not assessed; (3) toxicity outcomes were not considered; and (4) the paper was a review article. The details of this selection process are summarized in Figure 1.

### Selection Criteria

Automated filters were applied during the search to narrow down relevant studies. The inclusion criteria were as follows: Studies published between 2010 and 2025; Full-text, open-access, original research articles; Published in the English language; and Investigating chromium (VI)-induced neurotoxicity in animal models, human subjects, or cell-based systems. The exclusion criteria were as follows: Review articles, editorials, or case reports; Studies not assessing neuronal tissues or brain regions; Studies not involving Cr(VI) exposure or focusing on non-neurological outcomes; and Reports involving mixed or confounding metal exposures (Pb, Hg, Cd, Ni) in which chromium-specific effects could not be clearly distinguished.

To minimize confounding, studies that involved co-exposure to other heavy metals such as lead (Pb), mercury (Hg), cadmium (Cd), or nickel (Ni) were carefully evaluated. Only those providing analytical distinction or reporting isolated Cr(VI)-specific findings were prioritized for inclusion, whereas studies lacking clear separation were excluded. After applying all filters, the number of articles

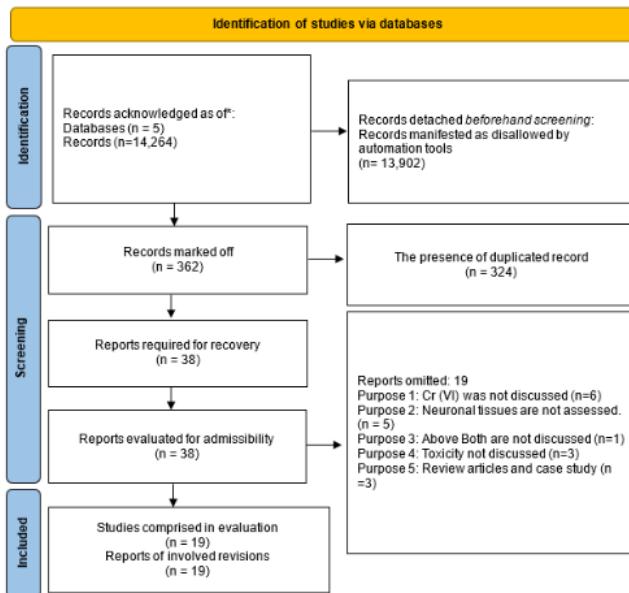


Figure 1. PRISMA Flow Diagram

was reduced to 362.

#### Data Extraction

Two independent reviewers conducted the screening and data extraction processes. Any discrepancies between reviewers were resolved through discussion and consensus to ensure accuracy and minimize selection bias. During data extraction, information on co-exposure to other metals and environmental contaminants was recorded to identify potential confounders, and studies isolating Cr(VI)-specific effects were highlighted for detailed review.

#### Quality Assessment

The methodological quality and risk of bias of the included studies were systematically evaluated. For animal studies, SYRCLE's Risk of Bias tool was applied to assess sequence generation, allocation concealment, blinding, incomplete outcome data, and selective reporting. For observational human studies, the Newcastle–Ottawa Scale (NOS) was used, evaluating selection, comparability, and outcome domains. Most animal studies demonstrated a low to moderate risk of bias, primarily due to limited reporting of randomization and blinding. Human cohort and case–control studies scored 7–8 out of 9 on the NOS, indicating good methodological quality with minor concerns related to exposure assessment and potential confounding by other metals. A summary of the quality assessment results for all included studies is presented in (Table 1).

#### Results

In this section, the authors briefly present the findings of the systematic literature review (SLR). The summarized results are provided separately in tabular form. Among the 19 studies included, 12 were animal-based and 7 were

human cohort studies. The detailed results are presented in (Table 2).

Table 3 summarizes experimental and animal studies investigating the neurotoxic effects of hexavalent chromium [Cr(VI)]. The findings indicate that Cr(VI) can cross the blood–brain barrier, leading to oxidative stress, neuroinflammation, apoptosis, and mitochondrial dysfunction. Evidence from studies on rodents, fish, and neuronal cell cultures demonstrates neuronal injury and behavioral impairments, whereas antioxidants such as salicylic acid and —coenzyme Q10 have shown potential neuroprotective effects.

Table 4 summarizes human observational and epidemiological studies examining the association between chromium and mixed-metal exposure and neurological outcomes.

Table 1. Summary of Quality and Risk-of-Bias Assessment of Included Studies

| Authors / Year                   | Assessment Tool | Score / Risk Level |
|----------------------------------|-----------------|--------------------|
| Ding et al. 2024 (20)            | SYRCLE          | Moderate           |
| Tang et al. 2024 (21)            | SYRCLE          | Moderate           |
| Salama et al. 2016 (22)          | SYRCLE          | Moderate           |
| Hegazy et al. 2021 (23)          | SYRCLE          | Low                |
| Kumari et al. 2014 (24)          | SYRCLE          | Moderate           |
| Hryntsova et al. 2022 (25)       | SYRCLE          | Moderate           |
| Chan et al. 2012 (26)            | SYRCLE          | Moderate           |
| Vielee et al. 2024 (27)          | SYRCLE          | Low                |
| Saleh et al. 2022 (28)           | SYRCLE          | Low                |
| Sedik et al. 2023 (29)           | SYRCLE          | Moderate           |
| Tripathi et al. 2022 (30)        | SYRCLE          | Moderate           |
| Zhu et al. 2019 (31)             | SYRCLE          | Low                |
| Baj et al. 2022 (32)             | NOS             | 8/9                |
| Rechtmann et al. 2020 (33)       | NOS             | 7/9                |
| Invernizzi et al. 2023 (34)      | NOS             | 8/9                |
| Viau et al. 2021 (35)            | NOS             | 7/9                |
| Heesterbeek et al. 2020 (36)     | NOS             | 7/9                |
| Parent et al. 2017 (37)          | NOS             | 7/9                |
| Figueroa-Romero et al. 2020 (38) | NOS             | 8/9                |

**Table 2.** Brief results of studies on neurological properties of Cr (VI)

| No | Authors / Year                   | Type of study                  | Place of study                                    | Relevance with current article   |
|----|----------------------------------|--------------------------------|---|--|
| 1  | Ding et al. 2024 (20)            | Investigational                | Guizhou Medical University                        | Evidence of Cr (VI) crossing BBB   |
| 2  | Tang et al. 2024 (21)            | Investigational                | Xiamen Medical College, China                     | Metabolic properties of Cr (VI) on Astrocytes                            |
| 3  | Salama et al. 2016 (22)          | Investigational                | National Research Centre, Egypt                   | Acute brain and lung injuries with intra-nasal exposure of Cr (VI)       |
| 4  | Hegazy et al. 2021 (23)          | Investigational                | National Research Centre, Egypt                   | Behavioral and neurotoxicological defect after intra-nasal exposure      |
| 5  | Kumari et al. 2014 (24)          | Investigational                | NEERI, Nagpur, India                              | Measurement of oxidative stress biomarker in Cr (VI) toxicity            |
| 6  | Hryntsova et al. 2022 (25)       | Investigational                | Department of Morphology, Ukarain                 | Oxidative stress in induced Cr (VI) noxiousness measured in Pineal gland |
| 7  | Chan et al. 2012 (26)            | Investigational                | The University of Hong Kong                       | Cr enhanced MRI of retina  |
| 8  | Vielee et al. 2024 (27)          | Investigational                | University of Louisville, Louisville              | Selective accumulation of Cr (VI) in hippocampus                         |
| 9  | Saleh et al. 2022 (28)           | Investigational Interventional | Ain Shams University, Egypt                       | SA attenuates the ROS production and protects against Cr (VI) toxicity   |
| 10 | Sedik et al. 2023 (29)           | Investigational Interventional | National Research Centre, Egypt                   | Neuroprotective effect of tangeretin (TNG)                               |
| 11 | Tripathi et al. 2022 (30)        | Investigational Interventional | ICMR, Ahmedabad, India                            | Beneficial properties of CoenzymeQ10, Biochanin A and Phloretin          |
| 12 | Zhu et al. 2019 (31)             | Investigational Interventional | College of Xinjiang Uyghur Medicine, China        | Protection by Aiweixin contrary to Cr (VI) toxicity                      |
| 13 | Baj et al. 2022 (32)             | Retrospective cohort study     | Medical University of Lublin, Poland              | Pathomechanism of neurodegeneration after multi element exposure         |
| 14 | Rechtman et al. 2020 (33)        | Observational                  | School of Medicine at Mount Sinai, NY, USA        | Multiple metal exposure and adolescents behavior                         |
| 15 | Invernizzi et al. 2023 (34)      | Observational                  | Icahn School of Medicine at Mount Sinai, New York | Multi metal exposure and functioning of brain networks in adolescents    |
| 16 | Viau et al. 2021 (35)            | Observational                  | Defense, Health, Environment, Centre Léon, France | Delayed nucleoschuttling of DSB by ATM protein                           |
| 17 | Heesterbeek et al. 2020 (36)     | Case control Observational     | Radboud University, Netherlands                   | Association of trace elements with age related macular degeneration      |
| 18 | Parent et al. 2017 (37)          | Observational                  | INTERPHONE study (7 participating countries)      | Association of glioma risk and occupational exposure of heavy metals     |
| 19 | Figueroa-Romero et al. 2020 (38) | Case control retrospective     | University of Michigan                            | Association of metal exposure and development of ALS                     |

Chromium was frequently co-detected with lead, copper, nickel, or manganese. Elevated chromium levels were correlated with impaired cognition, altered brain connectivity, and increased oxidative stress markers. Postmortem analyses revealed neural metal accumulation, whereas cohort studies reported variable findings, emphasizing the need for larger, well-controlled longitudinal investigations.

**Table 5** provides a comparative overview of the key neurotoxic mechanisms of chromium (VI) in relation to other major heavy metals, lead (Pb), mercury (Hg), and cadmium (Cd). While all induce oxidative stress and neuroinflammation, Cr(VI) uniquely crosses the blood-brain barrier via anion channels and preferentially accumulates in the hippocampus and hypothalamus. In contrast, Pb and Hg cause more pronounced disruptions in neuronal signaling and mitochondrial function, whereas Cd primarily interferes with DNA repair processes. The table also highlights distinct antioxidant- and chelation-based protective strategies relevant to each metal.

## Discussion

This section elaborates and synthesizes the findings of the studies included in the final review. The discussion

is organized into three subsections corresponding to the research objectives and emphasizes the mechanistic pathways of oxidative stress, inflammation, apoptosis, and mitochondrial dysfunction underlying Cr(VI)-induced neurotoxicity.

### **The findings of Experimental studies - On animals**

Collectively, animal studies identify oxidative stress as the primary mechanism by which Cr(VI) induces neurotoxicity, with inflammation, apoptosis, and mitochondrial dysfunction acting as interconnected pathways contributing to neuronal injury. Cr(VI) has been shown to cross the blood-brain barrier (BBB), accumulate in specific brain regions, and cause dose- and time-dependent oxidative damage. Experimental methods of Cr(VI) administration included intraperitoneal, intranasal, and oral routes, while assessment techniques such as immunohistochemistry, electron microscopy, and biochemical analysis were used to evaluate neurotoxicity. Salma A. et al. reported that intranasal administration of potassium dichromate (PDC) in rats produced a dose-dependent increase in chromium accumulation in the brain, with up to 46% of the administered dose reaching the brain at 2 mg/kg. Intraperitoneal administration at 15 mg/kg resulted in 36%

**Table 3.** Key Points of Animal Studies Examining Chromium (VI)-Induced Neurotoxicity and Protective Interventions

| Authors / Year             | Samples Population   | Chromium doses  | Tissue / Biomarkers studied   | Key findings   |
|----------------------------|--|---|---|--|
| Ding et al. 2024 (20)      | 57 mice  | 6 mg/kg (intraperitoneal) for 14 and 28 days            | Several brain regions   | Hypothalamus was identified as area of entry   |
| Tang et al. 2024 (21)      | Rat brain astrocyte culture                                  | 0–16 $\mu$ M Cr(VI) for 24 h                            | Brain astrocytes  | Mitochondrial disruption and apoptosis   |
| Salama et al. 2016 (22)    | 30 male Wistar rats  | 0.5–2 mg/kg (intranasal) and 15 mg/kg (intraperitoneal) | Overall brain   | 42% of instilled dose intranasally was able to reach to the brain                                  |
| Hegazy et al. 2021 (23)    | 32 male albino Wistar rats                                   | 0.125–0.5 mg/kg for 2–8 weeks                           | Astrocytes and Oligodendroglia  | Major toxic effects have come on locomotor and cognitive functions                                 |
| Kumari et al. 2014 (24)    | Lbeo Rohita, freshwater fish                                 | 48.3 ppm Cr(VI) for up to 15 days                       | Liver, Muscle, Gills, and Brain Oxidative stress                                  | Catalase, SOD, Glutathione Reductase activities increased  |
| Hryntsova et al. 2022 (25) | 24 white sexually mature male rats                           | 0.1 mg/L $K_2CrO_7$ in water (90 days)                  | Pineal gland extraction and GPX-1 estimation                                      | Abnormal morphological changes in cells and elevated levels of GPX-1                               |
| Chan et al. 2012 (26)      | 36 Adult Sprague-Dawley rats                                 | 1–100 mM PDC (intravitreal)                             | CrMRI was accomplished at 1-day postinjection upto two weeks                      | The iris showed a posology-dependent growth in T1-abnormal high hypertensity                       |
| Vielee et al. 2024 (27)    | 162 man and woman Sprague-Dawley scale of diverse age groups | 0.05–0.1 mg/L Cr(VI) in drinking water for 90 days      | Brain stem, cerebral cortex, cerebellum, hippocampus, hypothalamus, and striatum  | Geriatric female hippocampus accumulates the largest amount of Cr (VI)                             |
| Saleh et al. 2022 (28)     | 40 Wistar male rats  | 10–200 mg/kg PDC $\pm$ SA                               | AChE, MAOA, Dopamine, 5-HT, NAD+, HSP70, caspase-3, protein reporting, DNA injury | SA significantly reduced the ROS production, DNA damage and neurotoxicity, increased S100B protein |
| Sedik et al. 2023 (29)     | 32 male adult Wistar rats                                    | 2 mg/kg PDC $\pm$ TNG 50–100 mg/kg                      | Bevioral changes, MDA, Nrf2 expression, TNF- $\alpha$ , IL-6, GSH, MDA            | Reduced toxic effects with pre-exposure of TNG   |
| Tripathi et al. 2022 (30)  | 40 Adult Swiss albino male mice                              | Cr(VI) 75 ppm $\pm$ antioxidants (CoQ10, BCA, PHL)      | GSH, SOD, ACEs activity, head DNA %, Nrf2 expression, LPO                         | Oxidative stress was found, reduced significantly after antioxidants                               |
| Zhu et al. 2019 (31)       | Nematodes  | 10 mM PDC $\pm$ 0.05–0.125 vol AWX                      | ROS manufacture   | AWX had the time dependent protection against Cr (VI) toxicity                                     |

Note: Units standardized to mg/kg (animal doses), ppm (aquatic/cellular exposures), and  $\mu$ M (in-vitro concentrations) for consistency.

**Table 4.** Key Points of Human Observational Studies Investigating Chromium (VI) and Multi-Metal Exposure-Related Neurotoxicity

| Authors / Year                   | Samples                          | Chromium or or metal exposure                        | Tissue / Biomarkers studied   | Key findings   |
|----------------------------------|----------------------------------|--|---|--|
| Baj et al. 2022 (32)             | 178 postmortem samples           | 51 elements incl. Cr (ICP-MS)                        | Inductively Coupled Plasma Mass Spectrometry (ICP-MS)   | Cr was originated as the part of element cluster deposited on optic pathway                                      |
| Rechtman et al. 2020 (33)        | 150 adolescents                  | Mn, Pb, Cu, Cr, Ni (ICP-MS)                          | Self-assessment scales for externalization behavior   | Pb, Cr, and Cu contributed most to associations between metals and externalizing symptoms.                       |
| Invernizzi et al. 2023 (34)      | 193 young adults                 | Mn, Pb, Cr, Cu (ICP-MS)                              | Resting-state practical MRIs scans for global and limited efficacy (global:GE; local:LE) in 111 brain areas           | Substantial undesirable relations among the metal blend and GE and LE  |
| Viau et al. 2021 (35)            | Cell cultures                    | Al, Cu, Zn, Ni, Cd, Pb, Cr, Fe ( $\leq$ 100 $\mu$ M) | Irradiation and Immunofluorescence  | Metal Induced delay in the nucleo-shuttling of ATM proteins which, consequently delay DNA recognition and repair |
| Heesterbeek et al. 2020 (36)     | 236 nAMD patients, 236 controls  | Trace metals via ICP-MS                              | Plasma concentrations of trace elements   | Significant alterations in trace metal points amid the patients with nAMD and controls.                          |
| Parent et al. 2017 (37)          | 2054 glioma cases, 5160 controls | Occupational exposure to Pb, Cd, Ni, Cr, Fe          | -   | No exposure -outcome relationship found  |
| Figueroa-Romero et al. 2020 (38) | 36 ALS patients, 31 controls     | Cr, Mn, Ni, Zn (ICP-MS)                              | Laser ablation-IC-PMS was managed to gain period sequence data of metal in teeth from autopsies or dental extractions | Metal levels were upraised in cases than in controls with early life exposure                                    |

Note: Observational studies revealed consistent associations between cumulative or mixed metal exposure and neuropsychiatric, neurodevelopmental, or neurodegenerative outcomes, underscoring Cr(VI) as a key component of the toxic metal burden.

brain distribution. Intranasally exposed rats exhibited reduced locomotor activity, elevated malondialdehyde

(MDA), and decreased glutathione (GSH) and catalase levels, indicating oxidative stress (22). The study pro-

**Table 5.** Comparative Overview of Neurotoxic Effects of Cr(VI) vs Other Heavy Metals

| Mechanism                             | Chromium (VI)   | Lead (Pb)   | Mercury (Hg)   | Cadmium (Cd)   |
|---------------------------------------|---|---|--|--|
| Blood-Brain Barrier (BBB) Interaction | Crosses BBB via anion channels; accumulates in hippocampus & hypothalamus | Disrupts BBB integrity; alters endothelial tight junctions  | Strong BBB permeability; binds thiols in endothelial cells   | Weak BBB penetration; accumulates mainly in choroid plexus |
| Primary Mechanism of Toxicity         | Oxidative stress, Cr(VI); Cr(III) reduction generating ROS                | Interferes with calcium signaling, induces oxidative stress | Binds to sulfhydryl groups causing mitochondrial dysfunction | Promotes oxidative stress and inhibits DNA repair enzymes  |
| Inflammatory Response                 | NF-κB activation, IL-1β, IL-6, TNF-α upregulation                         | Activates microglia and astrocytes                          | Induces pro-inflammatory cytokine release                    | Elevates TNF-α, IL-8, and metallothionein expression       |
| Neuronal Effects                      | Apoptosis, mitochondrial injury, synaptic dysfunction                     | Axonal degeneration, cognitive impairment                   | Neuronal necrosis, tremors, visual and motor dysfunction     | Synaptic loss, cognitive decline                           |
| Target Brain Regions                  | Hippocampus, hypothalamus, pineal gland                                   | Cortex, hippocampus, cerebellum                             | Cerebellum, occipital cortex                                 | Hippocampus, cerebellum                                    |
| Protective Strategies                 | Antioxidants (CoQ10, TNG, SA, AWX) activate Nrf2 pathway                  | Chelation (EDTA), antioxidant support                       | Selenium, N-acetylcysteine                                   | Zinc supplementation, antioxidants                         |
| Mechanism                             | Chromium (VI)   | Lead (Pb)   | Mercury (Hg)   | Cadmium (Cd)   |

posed intranasal exposure as a suitable experimental model for simulating occupational Cr(VI) exposure in industries such as chromium plating and steel manufacturing. These findings support the hypothesis that oxidative imbalance initiates cellular damage, which is subsequently amplified by inflammatory cascades and apoptotic responses. Hegazy R. et al. examined repeated intranasal PDC administration in rats—(0.125–0.5 mg/kg/day, five days per week) and reported neuronal loss, astrocyte proliferation, and cognitive and locomotor impairments, further supporting occupational exposure relevance (23). Ding J. et al. and Tang H. et al. established a mechanistic sequence beginning with BBB disruption, followed by microglial activation, and culminating in mitochondrial dysfunction and neuronal apoptosis. These effects were mediated by increased expression of inflammatory markers (NF-κB, IL-1β, IL-6, TNF-α) and decreased antioxidant capacity (Nrf2, Nqo-1, Ho-1, 8-OHDG) (23).

Comparatively, studies such as those by Vielee S.T. et al. and Hryntsova N. et al. emphasized region-specific vulnerability, with the hippocampus, hypothalamus, and pineal gland being particularly susceptible due to their high metabolic activity and distinctive vascular characteristics (25, 27). These studies also reported sex- and age-dependent variations, with older female rats exhibiting greater Cr(VI) accumulation (27). Moreover, mitochondrial dysregulation, metal dyshomeostasis (Fe, Cu), and apoptosis consistently emerged as common patterns across different models, linking Cr(VI) exposure to neuronal degeneration and aging-like phenotypes. In one study, rats were exposed to Cr(VI) through drinking water for 90 days at doses of 0.05 and 0.1 mg/L, with the highest accumulation observed in geriatric females. Differences in gastric characteristics and Cr(VI) absorption related to age and sex were suggested as possible contributing factors. The study also demonstrated dyshomeostasis of essential metals (Fe, Cu, Se, Mn, Co, Mg), with gerontogenic effects observed in exposed young female rats (27). Dysregulation of essential elements such as iron (Fe) and copper (Cu) contributes to age-related neurological alterations.

Excess Fe is associated with neurodegenerative diseases, whereas Cu imbalance disrupts mitochondrial function and contributes to various disorders (39). Kumari et al. exposed Labeo rohita fish to Cr(VI), revealing increased oxidative stress markers in the liver, muscle, gills, and brain (24). Tang H. et al. analyzed Cr(VI)-treated rat astrocytes and found elevated ROS and 8-OHDG—levels, along with disrupted mitochondrial membranes, indicating oxidative stress and DNA damage. They also observed blood-brain barrier (BBB) disruption, altered sphingolipid metabolism, and abnormalities in the methionine-cysteine cycle, all associated with apoptosis and oxidative injury (21). Similarly, Hryntsova N. et al. reported oxidative stress in the pineal gland after 90 days of Cr(VI) exposure, accompanied by strong antioxidant protection (GPX-1). Morphological changes, including vascular damage and astrocyte activation, were evident; however, BBB disruption was not required due to the pineal gland's anatomical location (25). Additionally, Chan K.C. et al. demonstrated dose-dependent chromium toxicity via intravitreal potassium dichromate injections. Radiological imaging revealed increased retinal Cr contrast, while histological analysis showed altered lipid distribution. The reduction of Cr(VI) to Cr(III) generates reactive oxygen species, leading to disrupted lipid metabolism and impaired axonal transport (26). Interventional animal studies have highlighted the therapeutic potential of antioxidants. Coenzyme Q10, Biochanin A, Phloretin, Tangeretin (TNG), Aiweixin (AWX), and Sodium Alginate (SA) were shown to reduce ROS generation, restore antioxidant enzyme activity, and attenuate apoptosis (28–31). Collectively, these findings reveal a consistent mechanistic pattern: Cr(VI)-induced oxidative stress activates inflammatory and apoptotic pathways, leading to neuronal damage that can be mitigated through antioxidant modulation of the Nrf2/HO-1/NQO1 signaling axis. Further supporting evidence was provided by Tripathi S. et al., who investigated the neuroprotective effects of Coenzyme Q10, Biochanin A, and Phloretin in 40 mice exposed to arsenic and Cr(VI). Antioxidant treatment reduced metal accumulation, improved oxidative stress

markers, and restored body weight in toxin-exposed groups (30). Similarly, Sedik et. al. demonstrated Tangretin's (TNG) protective effects against Cr (VI)-induced brain injury by enhancing Nrf2 signaling and reducing inflammation and apoptosis in rats (29). Zhu B. et. al. showed that Aiweixin (AWX) decreased ROS production, with moderate doses providing significant protection without observable toxicity (31).

To investigate the neuroprotective action of Sodium Alginate (SA), Saleh E. M. et. al. examined its effects against Cr (VI)-induced neurotoxicity, focusing on oxidative stress, apoptosis, and neural damage. SA, known for its antifungal, antidiabetic, antioxidant, and anti-inflammatory properties, significantly reduced oxidative stress, apoptosis, and cellular injury in treated rats (28). Despite these promising findings, a knowledge gap persists due to several limitations in existing studies. One study based its conclusions on short-term Cr (VI) exposure, which may not accurately reflect the neurotoxicity resulting from chronic environmental exposure (23). Small sample sizes were also reported as a limitation (21). Moreover, most studies primarily focused on Cr (VI) accumulation and biomarker assessments, while functional evaluations—such as behavioral alterations and sensori-motor performance—were not addressed (20, 22). Furthermore, emerging research connects Cr(VI)-induced neurotoxicity to broader “toxic aging” concepts. Wise et al. (2024) introduced the “Toxic Aging Coin”, illustrating how environmental toxicants, including metals, accelerate aging through oxidative stress, inflammation, and cellular senescence. This perspective supports the present review’s findings that oxidative and inflammatory cascades mediate both neurotoxicity and premature neurodegeneration (40). In addition, Cao et al. (2020) demonstrated that taxifolin mitigates Cr(VI)-induced endothelial dysfunction and inflammation in HUVEC and THP-1 cells by suppressing MAPK and NF-κB activation, indicating therapeutic potential through antioxidant and anti-apoptotic mechanisms (41). Similarly, Dhande and Pansare (2024) reported that extracts of Tridax procumbens significantly reversed rotenone-induced oxidative and behavioral deficits in zebrafish and fruit fly models, supporting the neuroprotective relevance of plant-derived antioxidants (42). Furthermore, several studies have shown comparable neuroprotective and antinociceptive effects of both synthetic and natural compounds, such as thiazolidine derivatives, in diabetic neuropathic pain models, emphasizing antioxidant and anti-inflammatory mechanisms consistent with those observed in the Cr(VI)-induced neurotoxicity pathway (43–45).

#### ***The findings of Observational studies on humans***

Human studies provide complementary yet less direct evidence, describing metal dyshomeostasis and mixed heavy metal exposures associated with neurotoxicity. Baj J. et al. reported increased Cr concentrations in the optic chiasma linked to neuropathological alterations (32), while Rechtman. et al. identified correlations between exposure to multiple metals (Mn, Pb, Cu, Cr, Ni) and

behavioral abnormalities in adolescents (33). However, it must be explicitly acknowledged that Cr(VI) rarely occurs in isolation in human studies; most observations involve co-exposure with other metals. This substantially limits the ability to attribute the observed effects specifically to hexavalent chromium. In the context of environmental monitoring, Oralbekova et al. (2021) proposed a mathematical data assimilation model to optimize real-time monitoring of atmospheric heavy metals, including chromium, in urban environments such as Almaty City (46). Similarly, Zhartybayeva et al. (2023) applied ARIMA-based mathematical modeling to predict water pollution trends in Kazakhstan’s Akmola region, emphasizing the ecological and public health implications of heavy metal contamination (47). These approaches highlight the critical role of mathematical modeling in the early detection and prevention of Cr(VI) exposure in human populations.

Invernizzi A. et al. reported that urinary Cr levels correlated with reduced brain network efficiency on fMRI, suggesting systemic neurotoxicity (34). Radiological studies have further highlighted the Radiation-Induced ATM Nucleo-Shuttling (RIANS) model, which links ATM protein translocation to radiation-induced cellular toxicity and genomic instability. Similarly, Viau M. et al. demonstrated that concurrent exposure to multiple metals (Al, Cu, Zn, Ni, Pd, Cd, Pb, Cr, Fe) delayed ATM nucleo-shuttling and impaired DNA repair, indicating that genotoxic stress arises primarily from cumulative metal burden rather than isolated Cr(VI) exposure (35). These findings underscore that direct evidence of Cr(VI)-specific effects in human studies remains limited, with most data derived from mixed-metal exposure scenarios.

In addition, Chen et al. (2019) provided molecular insights into Cr(VI)-induced carcinogenesis, identifying both genetic and epigenetic mechanisms—such as histone modification and miRNA dysregulation—that may also contribute to neural genomic instability. Moreover, Al-Hussaniy et al. (2022) demonstrated that Panax ginseng protects against chemotherapy-induced cardiotoxicity and oxidative stress through caspase-mediated apoptosis modulation, reinforcing the role of antioxidant phytochemicals in mitigating toxicant-induced organ injury, a mechanism relevant to Cr(VI)-associated neuroprotection (49). Singh et al. (2025) further confirmed the antioxidant potential of several traditional medicinal plants, emphasizing that phytochemical screening through antioxidant assays can guide future preclinical and clinical investigations in toxicology and neuroprotection (50). Heesterbeek T.J. et al. reported lower chromium concentrations in individuals with neovascular age-related macular degeneration (nAMD) compared with healthy controls, suggesting a potential protective effect of chromium against oxidative stress. The study also associated cadmium levels with smoking and barium levels with antihypertensive drug use (36). Regarding metal exposure and cancer risk, several metals are classified as carcinogenic to humans, while inorganic lead is categorized as a probable carcinogen. To assess whether occupational exposure to these metals increases glioma risk, Parent M.E. et al. conducted a case-control study including 2,054 glioma pa-

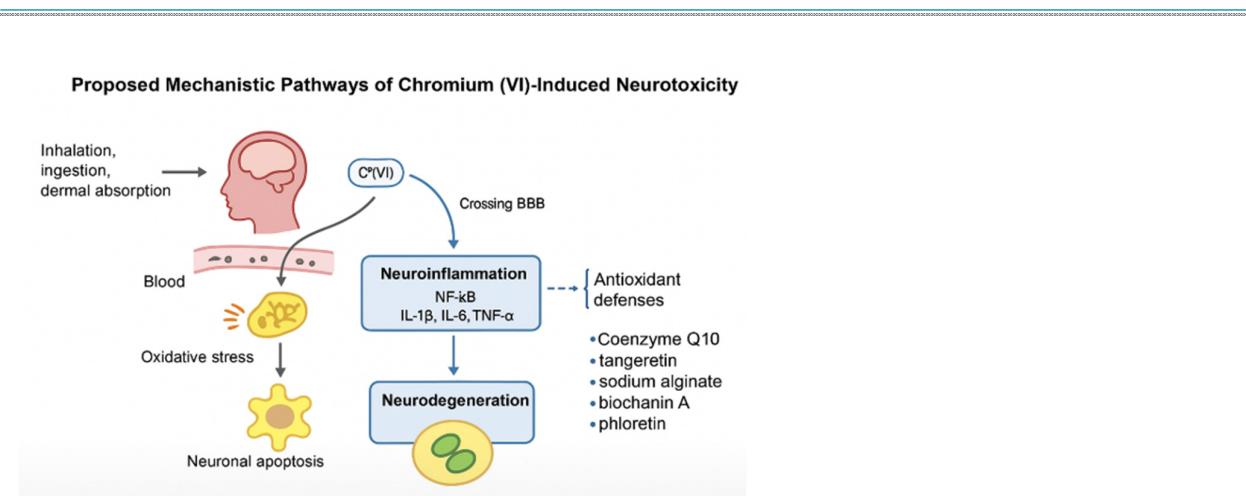


Figure 2. Proposed Mechanistic Pathways of Chromium (VI)-Induced Neurotoxicity

tients and 5,160 controls. No clear association was observed between exposure to lead, cadmium, nickel, chromium, or iron and glioma risk, despite exposure being assessed using the Finnish Job Exposure Matrix (FINJEM). The authors noted that a limited sample size reduced statistical power (37). Lee C. et.al. published a case report describing heavy metal toxicity in a parkinsonism patient with a metal-on-metal hip implant. MRI was used to visualize potential heavy metal (Cr and Co) deposition in the basal ganglia, which was associated with disease progression (51). These findings highlight the importance of careful decision-making when selecting prosthetic materials. However, larger case-control studies are needed to better elucidate the relationship between prosthetic-associated metal exposure and neurotoxicity. Identifying risk factors and developing radiological markers to assess Cr(VI) toxicity represent key research priorities for human cohort studies. Limitations noted in previous investigations include small sample sizes (35), subjective bias in self-reported questionnaires (28), an exclusive focus on linear associations while neglecting non-linearity, and a lack of systematic measurement approaches (34). The scarcity of consistent Cr(VI)-specific data, combined with small cohorts and confounding co-exposures (Pb, Cd, Ni, Fe), further restricts causal inference. Nevertheless, these studies collectively support chromium's contribution to oxidative and inflammatory neurotoxicity in humans, aligning with mechanisms established in animal models. Additionally, Mazakova et al. (2024) emphasized the growing role of mathematical modeling in toxicological and pharmacological research, demonstrating that such approaches can optimize pharmacokinetic and pharmacodynamic models, improve dose prediction, and enhance understanding of complex biological interactions (52). Incorporating these modeling frameworks could strengthen future Cr(VI) neurotoxicity research by improving exposure quantification, mechanistic prediction, and therapeutic optimization. Figure 2 illustrates the plausible mechanisms underlying Cr(VI)-induced neurotoxicity in the brain. Following inhalation, ingestion, or dermal absorption,

Cr(VI) enters systemic circulation and crosses the blood-brain barrier (BBB) via anion channels. Within neural cells, Cr(VI) undergoes stepwise intracellular reduction to Cr(V), Cr(IV), and Cr(III), generating excessive reactive oxygen species (ROS). Elevated ROS levels trigger oxidative stress, leading to lipid peroxidation, DNA damage, and mitochondrial dysfunction. Subsequent activation of NF-κB and associated inflammatory pathways promotes the release of pro-inflammatory cytokines (IL-1β, IL-6, TNF-α), resulting in neuroinflammation and microglial activation. Persistent oxidative and inflammatory stress further induce neuronal apoptosis through caspase activation, contributing to neurodegeneration in vulnerable brain regions such as the hippocampus and hypothalamus. Protective mechanisms include the activation of endogenous antioxidant defense pathways, particularly the Nrf2/HO-1/NQO1 signaling cascade, as well as the administration of exogenous antioxidants such as Coenzyme Q10, tangeretin, sodium alginate, biochanin A, and phloretin, which collectively mitigate oxidative damage and neuronal loss.

#### Integrative synthesis and clinical implications

Synthesizing evidence from both animal and human studies, Cr(VI)-induced neurotoxicity follows a multifactorial cascade involving oxidative stress, neuroinflammation, mitochondrial dysfunction, and apoptosis. This interconnected mechanistic sequence disrupts neural homeostasis, particularly in brain regions such as the hippocampus and hypothalamus. Antioxidant-based interventions that enhance Nrf2 signaling and suppress ROS generation consistently demonstrate neuroprotective effects, highlighting their potential translational relevance to clinical practice. Clinically, these findings emphasize the importance of proactive screening and preventive measures in chromium-exposed populations. Regular neurological assessments, monitoring of oxidative stress biomarkers, and dietary antioxidant supplementation may help mitigate early neurotoxic effects. Moreover, industrial and occupa-

tional health protocols should prioritize exposure surveillance, worker education, and safety compliance to minimize chromium-related risks. In summary, while animal studies provide well-defined mechanistic insights, human evidence remains constrained by co-exposure to multiple metals and methodological variability. Future longitudinal, exposure-controlled studies are essential to isolate Cr(VI)-specific effects and to evaluate targeted therapeutic strategies aimed at modulating oxidative and inflammatory pathways.

#### Limitations

This systematic review has several limitations. First, the small sample sizes of many included experimental and clinical studies limit the generalizability of the findings. Second, most data were derived from cross-sectional or short-term exposure studies, lacking longitudinal evaluation of cumulative effects. Third, only English-language and open-access studies were included, which may introduce selection and publication bias. Finally, variability in chromium measurement techniques and frequent co-exposure to other metals in human studies may confound the attribution of observed effects solely to Cr(VI).

#### Future Directions

##### Entry into the Brain

Future research should investigate the mechanisms by which Cr(VI) enters the brain, either through the blood-brain barrier (BBB) or via regions outside it and how its entry route, mode of administration, and target brain region influence neurotoxicity.

##### Factors affecting Neurotoxicity

Age and sex appear to modulate Cr(VI) neurotoxicity, with early-life exposure potentially increasing susceptibility to neurodegenerative changes. Cr(VI) may act as a gerontogen, disrupting metal homeostasis in a sex-dependent manner.

##### Sites of Neurotoxicity

Cr(VI) tends to accumulate in specific brain regions, particularly the hippocampus, where it affects multiple neuronal and glial cell types, leading to functional impairment.

##### Pathophysiological mechanisms

Cr(VI) induces oxidative stress, apoptosis, and neuroinflammation. Future studies should further explore its epigenetic effects and mitochondrial dysfunction, as well as their contributions to the progression of neurodegenerative processes.

#### Conclusion

The literature indicates that heavy metal ions such as Cr(VI), released from various industrial processes, are recognized as significant environmental toxins. The current systematic review summarizes recent research on Cr(VI)-induced oxidative stress and neuronal damage.

Animal-based experimental studies primarily elucidate the pathophysiological mechanisms of Cr(VI) toxicity in the brain and peripheral nervous system, whereas human cohort studies highlight associations and risk factors related to neurodegenerative and neurobehavioral disorders. Interventional studies predominantly conducted in animal models—have demonstrated the therapeutic potential of anti-oxidants in preventing and alleviating Cr(VI)-induced neurotoxicity. However, studies involving large cohorts of individuals with neurological disorders remain limited, representing a clear research gap in the existing literature. Future research should focus on population-based case-control models exploring the relationship between Cr(VI) exposure and diverse neurological conditions. Such investigations are essential to develop effective preventive strategies and to optimize therapeutic dosing for mitigating Cr(VI)-related neurotoxicity.

#### Authors' Contributions

MI: Conceptualization, project administration, funding acquisition, and supervision.

NA: Methodology, literature search (PubMed, Scopus, Web of Science), data extraction, and writing – original draft preparation.

SR: Data curation, quality assessment of included studies, and critical review of the manuscript.

YI: Formal analysis, methodology support, and validation of standardized units.

SS: Quality and risk-of-bias assessment using SYRCLE and NOS tools.

GS: Validation of results, visualization of mechanistic pathways, and final editing.

All authors have read and agreed to the published version of the manuscript.

#### Ethical Considerations

Not applicable.

#### Acknowledgment

Not applicable.

#### Conflict of Interests

The authors declare that they have no competing interests.

#### References

1. DesMarias TL, Costa M. Mechanisms of chromium-induced toxicity. *Curr Opin Toxicol*. 2019;14:1–7.
2. Moreira L de PD, Gomes JVP, Mattar JB, Chaves LO, Martino HSD. Potential of trace elements as supplements for the metabolic control of Type 2 Diabetes Mellitus: A systematic review. *J Funct Foods*. 2019;57:217–27.
3. Gelsanda CI, Marchianti ACN, Nurdian Y. The Relationship Between Well Water Quality and Cognitive Function in Well Water Users. *Futurity Medicine*. 2024;3:27–37.
4. Shridhar AM, Prashant SA, Rishi SA, Sunil AP, Vilas MA. Characterization of D-Limonene and its applications as antimicrobial antioxidant and enhancing of drinking water quality. *Res J Pharm Technol*. 2024;17:4591–6.
5. Aarabi S, Chauiyakh O, Ninich O, El Fahime E, Kettani K, Et-

Tahir A. Accumulation of heavy metals in water and sediments Moroccan Atlantics estuary. *Res J Pharm Technol.* 2023;16:5637-42.

6. Bhardwaj CK, Talniya NC, Mishra A, Singh VP, Rafi S. Synthesis, Spectral, Chromatographic and Antimicrobial Studies of Transition Metal Complexes of Chromium (III) and Cobalt (II) Ions with Alprazolam Drug. *Res J Pharm Technol.* 2024;17:3213-7.

7. Li MY, Shi YC, Xu WX, Zhao L, Zhang AZ. Exploring Cr(VI)-induced blood-brain barrier injury and neurotoxicity in zebrafish and snakehead fish, and inhibiting toxic effects of astaxanthin. *Environmental Pollution.* 2024;355:124280.

8. Zhu Y, Yan J, Xia L, Zhang X, Luo L. Mechanisms of Cr(VI) reduction by *Bacillus* sp. CRB-1, a novel Cr(VI)-reducing bacterium isolated from tannery activated sludge. *Ecotoxicol Environ Saf.* 2019;186:109792.

9. Singh V, Singh N, Verma M, Kamal R, Tiwari R, Sanjay Chivate M, et al. Hexavalent-Chromium-Induced Oxidative Stress and the Protective Role of Antioxidants against Cellular Toxicity. *Antioxidants.* 2022;11:2375 2022;11:2375.

10. Zhigalenok Y, Tazhibayeva A, Kokhmetova S, Starodubtseva A, Kan T, Isbergenova D, et al. Hexavalent chromium at the crossroads of science, environment and public health. *RSC Adv.* 2025;15:21439-64.

11. Gawdi R, Shumway KR, Emmady PD. Physiology, Blood Brain Barrier. *StatPearls* 2023.

12. Zheng W, Gherzi-Egea JF. ToxPoint: Brain Barrier Systems Play No Small Roles in Toxicant-induced Brain Disorders. *Toxicological Sciences.* 2020;175:147-8.

13. Harrison-Brown M, Scholes C, Field C, McQuilty R, Farah SB, Nizam I, et al. Limited penetration of cobalt and chromium ions into the cerebrospinal fluid following metal on metal arthroplasty: a cross-sectional analysis. *Clin Toxicol.* 2020;58:233-40.

14. Li MY, Shi YC, Xu WX, Zhao L, Zhang AZ. Exploring Cr(VI)-induced blood-brain barrier injury and neurotoxicity in zebrafish and snakehead fish, and inhibiting toxic effects of astaxanthin. *Environmental Pollution.* 2024;355:124280.

15. Bertollo AG, Santos CF, Bagatini MD, Ignacio ZM. Hypothalamus-pituitary-adrenal and gut-brain axes in biological interaction pathway of the depression. *Front Neurosci.* 2025;19:1541075.

16. Wise JP, Young JL, Cai J, Cai L. Current understanding of hexavalent chromium [Cr(VI)] neurotoxicity and new perspectives. *Environ Int.* 2022;158:106877.

17. Rahayu S, Rohmatika AU, Islamatasya U, Susilo RJK, Hayaza S, Santoso D, et al. The Potential of *Ganoderma applanatum* Polysaccharides Extracts on Apoptosis, Necrosis, TNF- $\alpha$ , and IL-6 in Diethylnitrosamine-Induced Mice. *Res J Pharm Technol.* 2025;18:809-14.

18. Salve PS, Pohankar AA, George S, Gadge JR. Alzheimer's Disease: A Study on Natural and Herbal Treatment. *Res J Pharm Technol.* 2025;18:898-906.

19. Naveen Kumari K, Jeyabalai S, Rajangam J, Gopinathan N, Ramakrishnan SR, Jayashankar Reddy V. Neuroprotective Potential of Total Extract of *Ulva Lactuca*: An In vitro study. *Res J Pharm Technol.* 2023;16:5948-53.

20. Ding J, Sun B, Gao Y, Zheng J, Liu C, Huang J, et al. Evidence for chromium crosses blood brain barrier from the hypothalamus in chromium mice model. *Ecotoxicol Environ Saf.* 2024;273:116179.

21. Tang H, Li K, Lin L, Wang W, Jian W. Study on the metabolic effects of hexavalent chromium [Cr (VI)] on rat astrocytes using un-targeted metabolomics. *Front Mol Biosci.* 2024;11:1372783.

22. Salama A, Hegazy R, Hassan A. Intranasal Chromium Induces Acute Brain and Lung Injuries in Rats: Assessment of Different Potential Hazardous Effects of Environmental and Occupational Exposure to Chromium and Introduction of a Novel Pharmacological and Toxicological Animal Model. *PLoS One.* 2016;11:e0168688.

23. Hegazy R, Mansour D, Salama A, Hassan A, Saleh D. Exposure to intranasal chromium triggers dose and time-dependent behavioral and neurotoxicological defects in rats. *Ecotoxicol Environ Saf.* 2021;216:112220.

24. Kumari K, Khare A, Dange S. The Applicability of Oxidative Stress Biomarkers in Assessing Chromium Induced Toxicity in the Fish *Labeo rohita*. *Biomed Res Int.* 2014;2014:782493.

25. Hryntsova N, Hodorová I, Mikhaylik J, Romanyuk A. A Response of the Pineal Gland in Sexually Mature Rats under Long-term Exposure to Heavy Metal Salts. *Prague Med Rep.* 2022;123:225-42.

26. Chan KC, Fan SJ, Zhou IY, Wu EX. In vivo chromium-enhanced MRI of the retina. *Magn Reson Med.* 2012;68:1202-10.

27. Vielee ST, Buchanan WJ, Roof SH, Kahloon R, Evans E, Isibor J, et al. Chromium Selectively Accumulates in the Rat Hippocampus after 90 Days of Exposure to Cr(VI) in Drinking Water and Induces Age- and Sex-Dependent Metal Dyshomeostasis. *Toxicics.* 2024;12:722.

28. Saleh EM, Hamdy GM, Hassan RE. Neuroprotective effect of sodium alginate against chromium-induced brain damage in rats. *PLoS One.* 2022;17:e0266898.

29. Sedik AA, Elgohary R, Sedik AA, Elgohary R. Neuroprotective effect of tangeretin against chromium-induced acute brain injury in rats: targeting Nrf2 signaling pathway, inflammatory mediators, and apoptosis. *Inflammopharmacology.* 2023 31:3 2023;31:1465-80.

30. Tripathi S, Fhatima S, Parmar D, Singh DP, Mishra SD, Mishra R, et al. Therapeutic effects of CoenzymeQ10, Biochanin A and Phloretin against arsenic and chromium induced oxidative stress in mouse (*Mus musculus*) brain. *3 Biotech.* 2022;12:5 2022;12:1-13.

31. Zhu B, Jo K, Yang P, Tohti J, Fei J, Abuduikerim K. Aiweixin, a Traditional Uyghur Medicinal Formula, Extends the Lifespan of *Caenorhabditis elegans*. *Evidence-Based Complementary and Alternative Medicine.* 2019;2019:3684601.

32. Baj J, Forma A, Kowalska B, Teresiński G, Buszewicz G, Majerek D, et al. Multi-Elemental Analysis of Human Optic Chiasm—A New Perspective to Reveal the Pathomechanism of Nerve Fibers' Degeneration. *Int J Environ Res Public Health.* 2022;19:4420.

33. Rechtman E, Curtin P, Papazaharias DM, Renzetti S, Cagna G, Peli M, et al. Sex-specific associations between co-exposure to multiple metals and visuospatial learning in early adolescence. *Translational Psychiatry.* 2020 10:1 2020;10:1-10.

34. Invernizzi A, Rechtman E, Oluyemi K, Renzetti S, Curtin P, Colicino E, et al. Topological network properties of resting-state functional connectivity patterns are associated with metal mixture exposure in adolescents. *Front Neurosci.* 2023;17:1098441.

35. Viau M, Sonzogni L, Ferlazzo ML, Berthel E, Pereira S, Bodgi L, et al. DNA double-strand breaks induced in human cells by twelve metallic species: Quantitative inter-comparisons and influence of the ATM protein. *Biomolecules.* 2021;11:1462.

36. Heesterbeek TJ, Rouhi-Parkouhi M, Church SJ, Lechanteur YT, Lorés-Motta L, Kouvatsos N, et al. Association of plasma trace element levels with neovascular age-related macular degeneration. *Exp Eye Res.* 2020;201:108324.

37. Parent ME, Turner MC, Lavoué J, Richard H, Figueroa J, Kincl L, et al. Lifetime occupational exposure to metals and welding fumes, and risk of glioma: a 7-country population-based case-control study. *Environmental Health.* 2017;16:1-10.

38. Figueroa-Romero C, Mikhail KA, Gennings C, Curtin P, Bello GA, Botero TM, et al. Early life metal dysregulation in amyotrophic lateral sclerosis. *Ann Clin Transl Neurol.* 2020;7:872-82.

39. Haywood S. Brain-Barrier Regulation, Metal (Cu, Fe) Dyshomeostasis, and Neurodegenerative Disorders in Man and Animals. *Inorganics.* 2019;7:108.

40. Wise RM, Wise JP, Andersen JK, Aschner M. Editorial: A Toxic Aging Coin perspective to investigate the intersection of toxicology and aging. *Front Toxicol.* 2024;6:1527706.

41. Cao X, Bi R, Hao J, Wang S, Huo Y, Demoz RM, et al. A study on the protective effects of taxifolin on human umbilical vein endothelial cells and THP-1 cells damaged by hexavalent

chromium: a probable mechanism for preventing cardiovascular disease induced by heavy metals. *Food Funct.* 2020;11:3851–9.

42. Dhande SR, Pansare N. Neuroprotective Activity Guided Fractionation of *Tridax procumbens* in Zebra Fish and Fruit Fly Model. *Res J Pharm Technol.* 2024;17:3141–50.

43. Kaur M, Kaur N, Muthuraman A, Kumar S. The Neuroprotective and Antinociceptive effect of Antidiabetic 3-(2-chlorophenyl)-4-imino-5-phenyl-2-(2-methoxyphenyl)-2H, 3H-[1,2,5] thiadiazolidin-1-oxide (CIPMTO) in Streptozotocin-induced diabetic neuropathic pain in rats: primary proof of concept. *Res J Pharm Technol.* 2022;15:5405–14.

44. Dombaanai B, Karabano R, Baig S. The Effect of Iron Citrate Synthesis On Cognitive Functions In Mice. *South East Eur J Public Health.* 2025:104–11.

45. Kusnir P, Baig S. The Neuropathy as an Adverse Effect of Chemotherapy Diminished After Administration of Dietary Supplement with Iron Citrate – Case Study. *Futurity Medicine.* 2024;3.

46. Oralbekova Z, Khassenova Z, Mynbayeva B, Zhartybayeva M, Iskakov K. Information system for monitoring of urban air pollution by heavy metals. *Indonesian Journal of Electrical Engineering and Computer Science.* 2021;22:1590–600.

47. Zhartybayeva M, Muntayev N, Tulegenova S, Oralbekova Z, Lamasheva Z, Iskakov K. Monitoring and Forecasting of Water Pollution by Heavy Metals. *IEEE Access.* 2023;11:1593–602.

48. Chen QY, Murphy A, Sun H, Costa M. Molecular and Epigenetic Mechanisms of Cr(VI)-induced Carcinogenesis. *Toxicol Appl Pharmacol.* 2019;377:114636.

49. Al-Hussaniy HA, Mohammed ZN, Alburghaif AH, Naji MA. Panax ginseng as Antioxidant and Anti-inflammatory to reduce the Cardiotoxicity of Doxorubicin on rat module. *Res J Pharm Technol.* 2022;15:4594–600.

50. Singh M, Kamal YT, Verma N, Mishra A, Sharma V, Ahmad S. Anti-Oxidant Potential of some Herbal Drugs: A Bioactivity Guiding approach for Chronic Diseases. *Res J Pharm Technol.* 2025;18:513–21.

51. Lee C, Hu M, Lee C, Hu M. Parkinsonism in a Patient With Metal on Metal Total Hip Replacement Related Elevated Serum Heavy Metal Levels. *Cureus.* 2021;13.

52. Mazakova A, Jomartova S, Mazakov T, Ziyatbekova G, Tursynbai A. Application of mathematical methods in pharmacology. *Journal of Interdisciplinary Mathematics.* 2024;27:1421–35.