



Correlations between Specific and Nonspecific Vaginal Immunity in Women with Breast Cancer in Kazakhstan

Ainur Amanzholkyzy¹, Gulaim Taskozhina², Farida Balmaganbetova^{1*}, Azhar Zhexenova², Roza Nurgaliyeva¹

Received: 16 Dec 2020

Published: 24 Dec 2021

Abstract

Background: The most common malignant tumor in women is breast cancer (BC). The ability of regulatory cells to inhibit cellular immune response as well as to participate in the modulation of antitumor immunity has attracted much interest of scientists. The purpose of this study was to assess the correlation between the specific and nonspecific vaginal immunity in women with BC.

Methods: This was an experimental study. The study involved 278 women, 174 of whom received chemotherapy. The sampling was performed using a universal probe. The qualitative and quantitative assessment of the vaginal microflora was done using the polymerase chain reaction method. Statistical processing of the analysis was performed using the Statistica 10.0 licensed software. The parameters of the immune status before and after chemotherapy were analyzed, and the correlation between the number of cells in the main populations of lymphocytes before and after chemotherapy was investigated.

Results: The study of the correlation between the number of cells of the main lymphocyte populations before and after chemotherapy showed an inhibition of B-lymphocytes (CD3-CD19+) in the study group, as the subpopulations of T-cytotoxic (CD4-CD8+) and CD3+HLA-DR+ (activated E-lymphocytes) were increased in both groups. Direct correlations were observed between local vaginal immunity and the immune status of the examined women in the study group between *Megasphaera* spp. and Enterobacteriaceae, with a certain population of immunocompetent cells.

Conclusion: It was concluded that impaired biocenosis and suppression of local immune responses in women with BC were the reason for the active involvement of the components of the immune system.

Keywords: Malignant Neoplasms, Immunoediting, Vaginal Immunity, Normal Flora, Obligate Anaerobes, Biocenosis

Conflicts of Interest: None declared

Funding: Grant funding of research and technical project No. 0118PK01065 for 2018-2020 by the Ministry of Science and Education of the Republic of Kazakhstan.

*This work has been published under CC BY-NC-SA 1.0 license.

Copyright© Iran University of Medical Sciences

Cite this article as: Amanzholkyzy A, Taskozhina G, Balmaganbetova F, Zhexenova A, Nurgaliyeva R. Correlations between Specific and Nonspecific Vaginal Immunity in Women with Breast Cancer in Kazakhstan. *Med J Islam Repub Iran.* 2021 (24 Dec);35:174. <https://doi.org/10.47176/mjiri.35.174>

Introduction

Breast cancer (BC) is the most common malignant tumor among women worldwide, as well as in Kazakhstan. In the structure of malignancies among women in economically developed countries, BC is the most common, significantly outscoring the proportion of other neoplasms

(1). Every year more than 2,000,000 women are diagnosed with BC worldwide (10%-18% of all malignant neoplasms) (2, 3). In the Republic of Kazakhstan, up to 4000 new cases of this oncological disease are diagnosed annually (4). In many malignant neoplasms, various significant

Corresponding author: Farida Balmaganbetova, balmaganbetova5485@kpi.com.de

¹ Department of Normal Physiology, West Kazakhstan Marat Ospanov Medical University, Republic of Kazakhstan

² Department of Pathological Physiology, West Kazakhstan Marat Ospanov Medical University, Aktobe, Republic of Kazakhstan

↑What is “already known” in this topic:

An integrated assessment of the vaginal microbiocenosis and local immunity in patients with cervical intraepithelial tumors was made, where it was proved that the development of cervical tumors is associated with genetically determined significant dysbiotic processes in the vagina with predominant participation of obligate anaerobes and local immune dysfunctions.

→What this article adds:

The present study assesses the correlation between specific and nonspecific vaginal immunity in women with BC.

immune disorders are observed: a decrease in T-lymphocytes and their subpopulations, number and functional activity of natural killer cells, et cetera. Various populations of cells play an important role in antitumor defense of the body, including both effector and suppressor cells. At present, much attention is paid to the study of regulatory cells, including CD4+ and CD8+ lymphocyte subpopulations as well as NKT cells (5). These cells are able to inhibit cellular immune response as well as to participate in the modulation of antitumor immunity (6-8).

The immune system's interaction with cancer is a delicate balance between immune activation and immune suppression. The dual nature of the interaction between the immune system and the tumor is currently viewed as a dynamic process of immunoediting. An important role in the immune response to a tumor is played by various innate and adaptive immunity cell populations: NK-, T-, NKT-cells, macrophages, and dendritic cells. These populations are heterogeneous and contain both cells with antitumor activity and regulatory (suppressor) cells that promote tumor progression (9-11).

The specific cellular response of the urogenital mucous membranes is formed according to the T cell and B cell humoral pathways. The T cell-mediated immune response is aimed at the destruction of intracellular pathogens and is mediated mainly by CD8+ T-lymphocytes, which are located in the stroma of the vagina, cervix, and uterus under the epithelial layer, and also scattered between epithelial cells (12, 13).

The influence of the immune status, endocrine factors, and some neoplastic processes and their combination on the formation of vaginal dysbiosis was discussed in a number of scientific works. Reproductive health of a woman depends on complex mechanisms of regulation and cooperation between the epithelium of the mucous membrane of the reproductive system, local microflora, and immune cells, which produce biologically active substances and hormonal regulation (14, 15).

The purpose of this study was to assess the correlation between specific and nonspecific vaginal immunity in women with BC.

Methods

This was an experimental study. Research work was performed in the city of Aktobe in the Republic of Kazakhstan at the Medical Center of West Kazakhstan Marat Ospanov Medical University during 2018-2020. The study involved 278 women with BC from Kazakhstan, with a mean age of 56.7 ± 11.1 years. The women included in the study were divided into 2 observation groups. The study group included 174 patients (62.5%) who received chemotherapy. The control group included 104 patients (37.4%) before chemotherapy. All examined women gave their informed consent to participate in the study.

The qualitative and quantitative composition of the vaginal flora in women with BC was analyzed using the PCR using the Femoflor set of reagents. Materials were collected from the posterior fornix of the vagina. The sampling was done using a universal probe, the working

part of which, containing the test material, was cut off or broken off and placed in a disposable Eppendorf tube with a preservative solution (transport medium). These tubes were subsequently delivered to the polymerase chain reaction (PCR) laboratory of the Scientific and Practical Center of the Marat Ospanov West Kazakhstan Medical University, where the qualitative and quantitative assessment of the vaginal microflora was performed using the PCR method based on deoxyribonucleic acid (DNA) amplification. The content of microorganisms was expressed as the decimal logarithm of the absolute DNA number (16, 17).

Immunological studies included the determination of lymphocyte subpopulations based on the levels of expression of lymphocyte membrane antigens using a set of monoclonal antibodies specific to differentiation antigens (CD3, CD4, CD8, CD16, CD20, CD25, and CD95). The samples were analyzed using the FacsCalibur flow cytometer (Becton Dickinson) and the CellQuest software. The CellQuest software of the FacsCalibur flow cytometer allows the analysis of up to 50,000 cells in 1 sample simultaneously for several parameters: forward light scattering, side light scattering, and multicolor fluorescence, and to perform a multifactor analysis of cell populations (18, 19).

Statistical processing of this analysis was carried out using the Statistica 10.0 licensed software using the nonparametric Spearman's coefficient to compare the studied groups. The nonparametric Spearman's coefficient was used for independent samples and to assess the relationship between ordinal and quantitative characteristics.

The Spearman rank-order correlation coefficient is a nonparametric measure of the strength and direction of association that exists between 2 variables measured on at least an ordinal scale. This test is used for either ordinal variables or for continuous data that have failed the assumptions necessary for conducting Pearson's product-moment correlation. Methods of descriptive statistics with the calculation of central tendencies and their range were used for quantitative variables. The results were expressed as medians and upper and lower quartiles. Hypothesis testing was used to determine the P value on different stages of research. The significance level for the test was set at $P \leq 0.001$.

Results

The analysis of these results are described in stages in both groups for all parameters of specific immunity, nonspecific immunity, and relationship with immune status (Table 1). The analysis of the immune status after chemotherapy showed that practically all mean levels of cellular immunity according to the reference values of CD3+, CD19-, CD4+, CD8-, IRI, CD3-HLA-DR+, NK(CD16+56+), and CD3+/CD16+56+ were within the physiological normal range in all patients; however, CD3-CD19+ was significantly decreased compared with the reference value in the study group ($P \leq 0.001$); CD4-CD8+ and CD3+HLA-DR+ were significantly increased compared with the reference value in both groups.

Table 1. Comparative Analysis (Spearman's correlation) of Specific Immunity in Both Groups After Chemotherapy

Markers	Mean Values		P Value
	Study Group	Control Group	
T-lymphocytes (CD3+CD19-)	71.7 [66.3;76.7]	71.4 [65.4;75.8]	0.055
B-lymphocytes (CD3-CD19+)	7.9 [3.7;12.2]	10.4 [7.8;13.7]	0.001
T-helpers (CD4+CD8+)	40.7 [32.4;45.0]	39.0 [32.1;43.7]	0.036
T-cytokines (CD4-CD8+)	31.7[25.8;36.8]	28.5[24.5;35.4]	0.008
IRI	1.3[0.9;1.6]	1.3[1.0;1.7]	0.074
CD3+HLA-DR+(activated T- lymphocytes)	8.7 [4.5;14.8]	6.1 [2.0;13.0]	0.003
CD3-HLA-DR+	13.4 [9.8;15.4]	13.6 [11.5;14.9]	0.045
NK(CD16+56+)-natural killers	11.6[8.0;16.8]	11.3[7.3;18.8]	0.053
T-killers (CD3+/CD16+CD56+)	5.8[3.3;9.0]	6.4[4.5;10.0]	0.003

Note: Me – median, 25% – lower quartile, 75% – upper quartile

The analysis of the vaginal nonspecific immunity showed the following: *Peptostreptococcus* spp., *Gardnerella vaginalis* prevailed quantitatively in both groups, which indicated bacterial vaginosis, while the representative of obligate anaerobes, *Enterobacteriaceae*, showed high values in the control group. The representative of the obligate anaerobes, *Lachnobacterium* spp., representatives of the mycoplasma group, *ureaplasma* (*urealyticum* + *parvum*) and *Mycoplasma genitalium* showed high values in the study group, which indicated the presence of bacterial vaginitis and mycoplasmosis or ureaplasmosis.

As shown in Figure 1, the correlation analysis in the study group showed that among the representatives of obligate anaerobes, *Megasphaera* spp. had a medium direct correlation with CD4-CD8+ and CD3+HLA-DR+(activated E-lymphocytes) ($r = 0.5$; $P \leq 0.001$); a medium inverse correlation with the ratio of T-helpers to T-suppressors (IRI), CD3-HLA-DR+ ($r = -0.5$; $P \leq 0.001$); negative weak correlations between *Lachnobacterium* spp. and T-cytotoxic (CD4-D8+), CD3+HLA-DR+(activated E-lymphocytes) ($r = -0.2$; $r = -0.3$; $P \leq 0.001$), and a positive correlation with IRI ($r = 0.3$; $P \leq 0.001$). The following correlations were observed between *Mobiluncus* spp. and T-helper (CD4+CD8-) – weak direct ($r = 0.3$; $P \leq 0.001$), and *Peptostreptococcus* spp. and T-killer CD3+/CD16+56+ – medium direct ($r = 0.4$; $P \leq 0.001$). The representative of the mycoplasma group, *Ureaplasma* (*urealyticum* + *parvum*), showed a negative weak

correlation with B-lymphocyte (CD3-CD19+) ($r = -0.3$; $P \leq 0.001$). Figure 2 shows the correlation in the study group between the only representative of the facultative anaerobes, *Enterobacteriaceae*, and CD3-HLA-DR+ – positive correlation ($r = 0.6$; $P \leq 0.001$).

The following correlation analysis in the control group showed a strong inverse correlation between the representatives of obligate anaerobes: as shown in Figure 3, the correlation between *Peptostreptococcus* spp. and CD3+HLA-DR+ (activated E-lymphocytes) ($r = -0.6$; $P \leq 0.001$). Figure 4 shows that the correlation between *Sneathia* spp. and T-cytotoxic (CD4-CD8+) is strong and positive, and between *Sneathia* spp. and NK (CD16+56+)

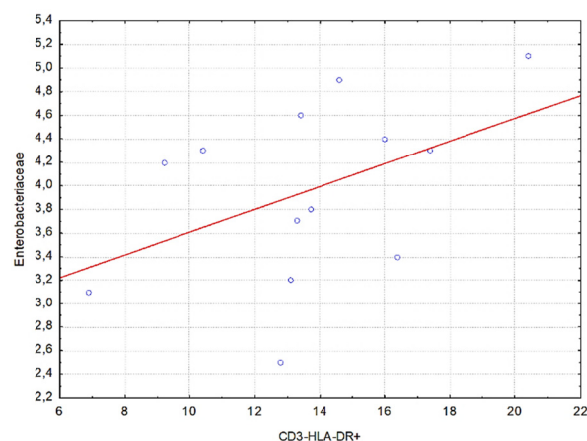


Fig. 2. Correlations Between Immunity and Enterobacteriaceae in the Study Group

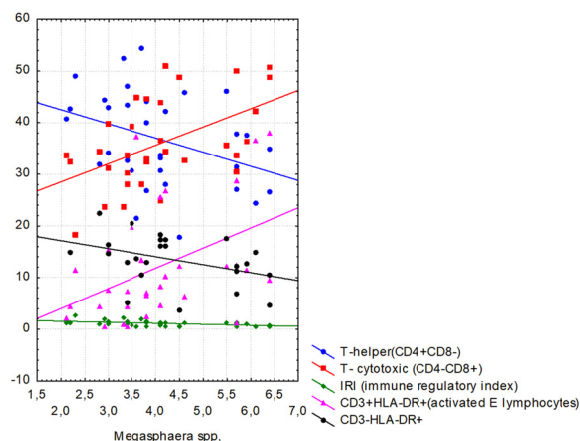


Fig. 1. Correlations Between Specific Immune Markers and *Megasphaera* spp. in the Study Group

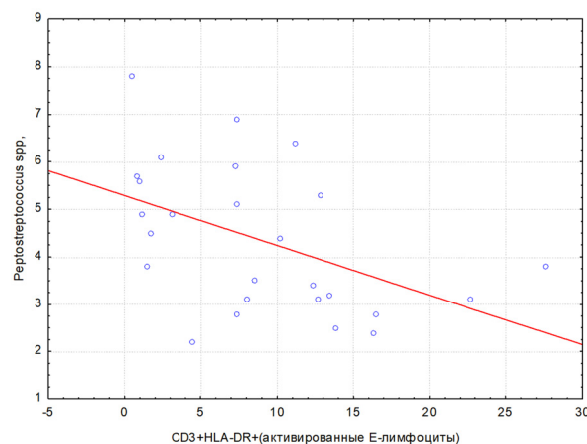


Fig. 3. Correlations Between Immune Markers and *Peptostreptococcus* spp. in the Control Group

natural killers is strong and negative ($r = -0.5$; $P \leq 0.001$). A weak but significant negative correlation was observed between *Gardnerella vaginalis* and CD3+HLA-DR+ (activated E-lymphocytes) ($r = -0.3$; $P \leq 0.001$).

Figure 5 also shows the correlation between the only representative of facultative anaerobes *Streptococcus* spp. and T-cytotoxic (CD4-CD8+), which was a significant direct medium correlation, while for IRI there was a significant inverse strong correlation ($r = 0.5$; $r = -0.6$; $P \leq 0.001$).

This was confirmed in our study by the fact that the development of an imbalanced immune response in tumors of the reproductive system affected the nonspecific vaginal immunity, with a predominance of representatives of obligate anaerobes with inflammation in the form of bacterial vaginosis (Table 2).

Discussion

The study of the relationship between specific immunity and nonspecific vaginal immunity markers showed a correlation between obligate anaerobes *Megasphaera* spp. and the main populations of lymphocytes, which were involved in the immune response to the infection and were directly involved in the destruction of infected cells (20). This was confirmed in our study and this fact indicated a significant decrease in the immune regulatory index (IRI) regarding a quantitative increase in *Megasphaera* spp. in the vaginal flora, which indicated the occurrence of bacterial vaginosis in patients in the study group. T-activated lymphocytes with the CD3+HLA-DR+ phenotype are the markers of late activation and immune hyperreactivity and a decrease in these parameters indicates a weakening of the body's defenses and the development of a hypimmune state (21). If T-helpers are up to 0.4×100 , the clinical picture can be transient and reversible. In our patients, it manifested itself as a correlation between these parameters and representatives of the vaginal biocenosis *Megasphaera* spp. and

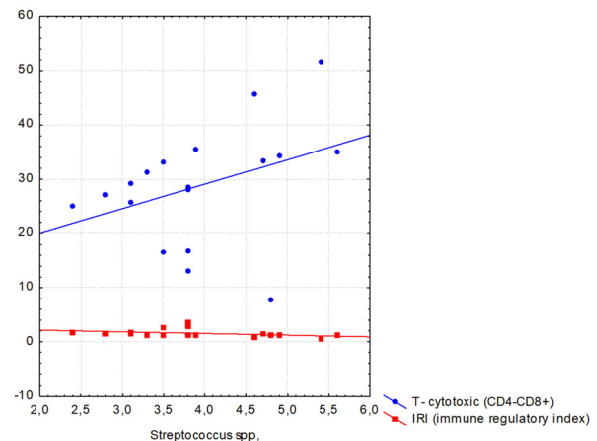


Fig. 4. Correlations Between Cellular Immunity and *Streptococcus* spp. in the Control Group.

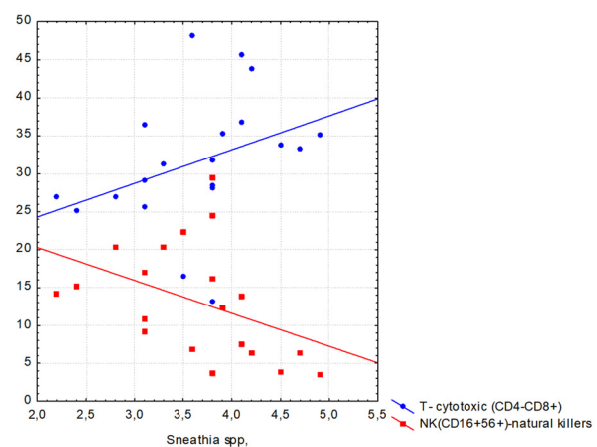


Fig. 5. Correlations Between Immune Markers and *Sneathia* spp. in the Control Group.

Enterobacteriaceae in women with BC, following long-term chemotherapy.

In the control group, the picture of the vaginal

Table 2. Correlation Between Immune Markers and Representatives of Vaginal Microflora in Both Groups With Breast Cancer

Immune markers	Representatives of Vaginal Microflora										
	Study group					Control Group					
	Enterobacteriaceae	Lachnabacterium spp.	Megasphaera spp.	Peptostreptococcus spp.	Mobiluncus spp.	Ureaplasma	Mycoplasma genitalium	Streptococcus spp.	Gardnerella vaginalis	Sneathia spp.	Peptostreptococcus spp.
T-cytotoxic (CD4-CD8+)		-0.2	0.5					0.5		0.5	
T-helper (CD4+CD8-)			-0.4		0.3						
CD3-HLA-DR+	0.6		-0.4								
CD3+HLA-DR+(activated E-lymphocytes)		-0.3	0.5						-0.3		-0.6
T-killers CD3+/CD16+56+				0.4							
B-lymphocytes (CD3-CD19+)			-0.4			-0.3					
NK(CD16+56+)-natural killers										-0.5	
IRI		0.3	-0.5		-0.2			-0.6			

Note: $P \leq 0.001$ was the significance level for all correlations.

biocenosis regarding this flora, such as *Peptostreptococcus* spp., *Sneathia* spp., and *Streptococcus* spp., changed with an increase in response to the inhibition of T-activated lymphocytes with the CD3+HLA-DR+ phenotype, IRI, and NK (CD16+56+)-natural killers at baseline in the examined women with BC before chemotherapy, and this indicates that in the acute phase of this pathology, the immune response is correspondingly reduced. Whereas in the control group, high values of the representative of obligate anaerobes, Enterobacteriaceae, had no correlations; and in the study group, the strong, direct correlation made it possible to consider it a late marker of cellular and nonspecific immunity. The results of this study in the control group in terms of correlation levels, among the presented vaginal biocenosis of *Peptostreptococcus* spp. and *Gardnerella vaginalis*, revealed an inverse correlation between CD3+HLA-DR+ and *Peptostreptococcus* spp., which may be considered an early marker of nonspecific immunity.

Based on the studies performed in Russia (Yekaterinburg authors) and in Europe, an integral assessment of the vaginal microbiocenosis and local immunity in patients with cervical intraepithelial tumors was made, where it was proved that the development of cervical tumors is associated with genetically determined significant dysbiotic processes in the vagina, with predominant participation of obligate anaerobes and local immune dysfunctions (22).

The authors from the Amsterdam Institute for Infection and Immunity state that bacterial vaginosis-associated *Megasphaera* spp. bacteria induced the maturation of dendritic cells and increased the levels of proinflammatory cytokines (23), which contradicts our results. This is because of the fact that *Megasphaera* spp. produced both direct and negative correlations with a subpopulation of T-lymphocytes in the study group.

The experimental study at the Chinese Research Institute of Hong Kong on mouse models with colorectal cancer showed that *Peptostreptococcus* spp. attached to the mucous membrane and accelerated the oncogenic response in vitro and in vivo (24). In our study, *Peptostreptococcus* spp. affected a subpopulation of T cells with inverse correlation (in vivo) (25).

Conclusion

The function of the immune system is to preserve and maintain beneficial bacteria in the vagina alive. The mechanisms of interactions between the immune system and normal microbiota remains unclear, but there is no doubt that the microbial biocenosis and the immunity of the vaginal mucosa function together. It is important to emphasize that the effects of microbial products are rather systemic than local. Neoplastic processes in the mammary glands are associated with genetically determined significant dysbiotic processes in the vagina with a predominant participation of obligate anaerobes and local immune dysfunctions. Among anaerobes, the most important are *Megasphaera* spp., *Streptococcus* spp., *Sneathia* spp., *Peptostreptococcus* spp. and *Enterobacterium* spp. The variety of leading pathogens

requires a comprehensive study of vaginal microbiocenosis and local immunity in patients with BC, with a correction of identified disorders to improve treatment outcomes. The components of the immune system (T-cytotoxic (CD4-CD8+) and CD3+HLA-DR+(activated E-lymphocytes)) are actively involved in maintaining nonspecific vaginal immunity, based on molecular genetic studies in women with BC, as manifested by an impaired biocenosis and depression of local immune responses.

The biocenosis of the vagina, together with the suppression of normal flora, indicates the prevalence of the representatives of obligate anaerobes: *Megasphaera* spp., Enterobacteriaceae, and *Peptostreptococcus* spp. in response to changes in the subpopulation of T cells associated with a decrease in immunobiological protection, creating conditions for the implementation of the pathogenic action of biocenosis, which further exacerbates the immunological failure. The presence of multiple correlations between biocenosis parameters and cellular immune status indicates the presence of a general immune response of the body and requires further in-depth study of these correlations.

Acknowledgment

This research was conducted in the city of Aktobe of the Republic of Kazakhstan at the Medical Centre of the Marat Ospanov West Kazakhstan Medical University, within the framework of grant funding of research and technical project No. 0118PK01065 for 2018-2020 by the Ministry of Science and Education of the Republic of Kazakhstan.

Conflict of Interests

The authors declare that they have no competing interests.

References

1. Kalieva G. Breast cancer: Targeted, safe, effective. *Kazakhstan Med J*. 2014;5(41):27–29.
2. Forman D, Bray F, Brewster DH, Gombe Mbalawa C, Kohler B, Piñeros M, et al. Cancer incidence in five continents. Vol. X. Lyon: International Agency for Research on Cancer. 2014.
3. Bilyalova ZA, Iginov NS, Turemuratova MA. Epidemiological aspects of breast cancer in Kazakhstan. *Young Sci*. 2011;1(24):257–261.
4. Wang K, Vella AT. Regulatory T cells and cancer: A two-sided story. *Immun Invest*. 2016;45(8):797–812.
5. Bower JE, Ganz PA, Aziz N, Fahey JL, Cole SW. T-cell homeostasis in breast cancer survivors with persistent fatigue. *J Nation Cancer Instit*. 2003;6/95(15):1165–8.
6. Paleev FN, Sanina NP, Makarkov AI, Mylov NM, Ostrovsky EI, Khishova NN. Immunological characteristics of inflammatory diseases of the myocardium of viral etiology. *Almanac Clin Med*. 2014;35:12–21.
7. Byrne A, Savas P, Sant S, Li R, Virassamy B, Luen SJ, et al. Tissue-resident memory T cells in breast cancer control and immunotherapy responses. *Nature Rev Clin Oncol*. 2020;17:341–348.
8. Sumransub N, Jirapongwattana N, Jamjuntra P, Thongchot S, Chiochansin T, Yenchitsomanus PT, et al. Breast cancer stem cell RNA-pulsed dendritic cells enhance tumor cell killing by effector T cells. *Oncol Lett*. 2020;19(3):2422–2430.
9. Grigorieva TA, Beznos OA, Tupitsyn NN, Vorotnikov IK, Selchuk VYu, Ryabchikov DA. Subpopulations of bone marrow lymphocytes

- in breast cancer patients. *Tumors Fem Reprod Sys.* 2015;2:52-55.
10. Korotkova OV, Zabolina TN, Skotarenko LV, Borunova A, Ocheeva NYu, Vorotnikov IK, et al. Subpopulations of peripheral blood lymphocytes in breast cancer patients. *Russ Biotherap J.* 2011;3(10):95-98.
 11. Avdeeva ZH, Soldatov AA, Kiselevsky MV, Bondarev VP, Merkulov VA. Antineoplastic drugs of monoclonal antibodies. *Med Immun.* 2017;19:157-158.
 12. Baxevanis CN, Fortis SP, Perez SA. The balance between breast cancer and the immune system: Challenges for prognosis and clinical benefit from immunotherapies. *Semin in Canc Biol.* 2019. <https://pubmed.ncbi.nlm.nih.gov/31881337/>
 13. Shamilov FA, Vishnevskaya YaV, Selchuk VYu, Pogodina EM, Zernov DI, Chkhikvadze NV, et al. Possibilities of studying subpopulations of intratumoral lymphocytes by flow cytometry on the material of core-biopsy of a tumor in patients with breast cancer. *Tumors Fem Reprod Sys.* 2012;3/4:29-33.
 14. Lebedeva OP, Kalutsky PV, Pakhomov SP, Churnosov MI, Karpov PA. Congenital immunity of the female reproductive tract and its hormonal regulation (mini-review). *Sci Stat.* 2009;12(67):25-30.
 15. Stakheeva MN, Eydenzon D, Slonimskaya EM, Kukharev YaV, Garbukov EYu, Babyshkina NN, et al. The relationship between the state of the immune system as an integrated whole and the clinical course of breast cancer. *Siberian J Oncol.* 2011;2(44):11-19.
 16. Beznos OA, Burov DA, Selchuk VYu, Vorotnikov IK, Timoshenko VV, Grigorieva TA, et al. Interrelation of intratumoral lymphocyte subpopulations with clinical and pathomorphological features of breast cancer. *Tumors Fem Reprod Sys.* 2016;2:13-17.
 17. Laniewski P, Barnes D, Goulder A, Cui H, Roe DJ, Chase DM, et al. Cervicovaginal immune signatures, HPV and microbiota composition in cervical carcinogenesis in non-Hispanic and Hispanic women. *Sci Rep.* 2018;15(8(1):7593.
 18. Long X, Wong ChCh, Tong L, Chu ESH, Szeto ChH, Go MYYY, et al. Peptostreptococcus anaerobius promotes colorectal carcinogenesis and modulates tumour immunity. *Nature Microbiol.* 2019;4(12):2319-2330.
 19. Skotarenko LV, Vorotnikov IK, Kadagidze ZG, Shamilov FA. Features of T-cell immunity in breast cancer. *Tumors of the Fem Reprod Sys.* 2011;4:24-27.
 20. Ablin RJ, Bhatti RA, Guinan PD, Khin W. Modulatory. Effects of oestrogen on immunological responsiveness. II. Suppression of tumour-associated immunity in patients with prostatic cancer. *Clin Exper Immunol.* 1979;38(1):83-91.
 21. Tabakov DV, Zabolina TN, Zakharova EN, Borunova AA, Korotkova OV, Chertkova AI, et al. Subpopulation balance of peripheral blood effector cells in cancer patients. *Immunology.* 2019;40(3):20-27.
 22. Lebedeva OP, Kalutsky PV, Pakhomov SP, Churnosov MI, Karpov PA. Congenital immunity of the female reproductive tract and its hormonal regulation (mini-review). *Sci Stat.* 2009;12(67):25-30.
 23. van Teijlingen NH, Helgers LC, Zijlstra-Willems EM, van Hamme JL, Ribeiro CMS, Strijbis K, et al. Vaginal dysbiosis associated-bacteria *Megasphaera elsdenii* and *Prevotella timonensis* induce immune activation via dendritic cells. *J Reprod Immunol.* 2020;138:103085.
 24. Sconocchia G, Eppenberger S, Spagnoli GC, Tomillo L, Droeser R, Caratelli S, et al. NK cells and T cells cooperate during the clinical course of colorectal cancer. *Oncoimmunology.* 2014;3/3(8):e952197.
 25. Herías MV, Midtvedt T, Hanson LA, Wold AE. Increased antibody production against gut-colonizing *Escherichia coli* in the presence of the anaerobic bacterium *Peptostreptococcus*. *Scand J Immunol.* 1998;48(3):277-282.